

# Light Microscopy, Super Resolution Microscopy and Correlative Light and Electron Microscopy

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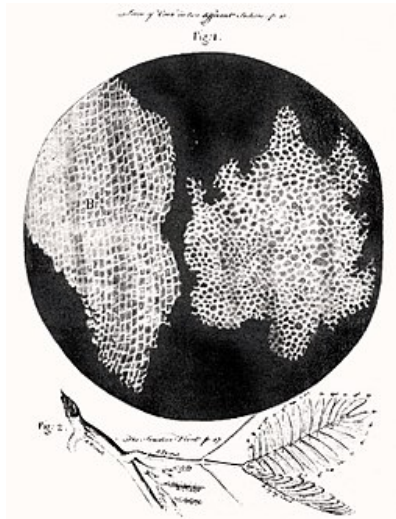
## Outline

- Introduction to light microscopy
- Fluorescence microscopy
- Super resolution microscopy
- Correlative light and electron microscopy

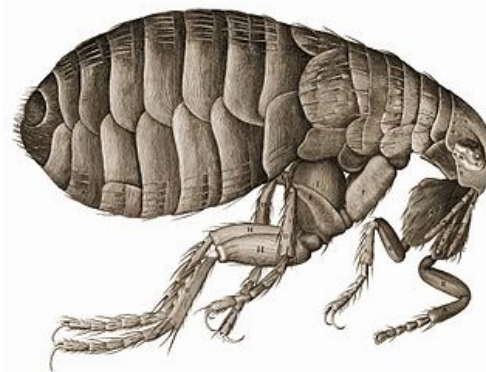
# Robert Hooke, “the father of microscopy” (1665)

In 1665 Hooke published *Micrographia*, a book describing observations made with microscopes and telescopes, as well as some original work in biology.

Hooke coined the term **cell** for describing biological organisms, the term being suggested by the resemblance of plant cells to cells of a honeycomb.



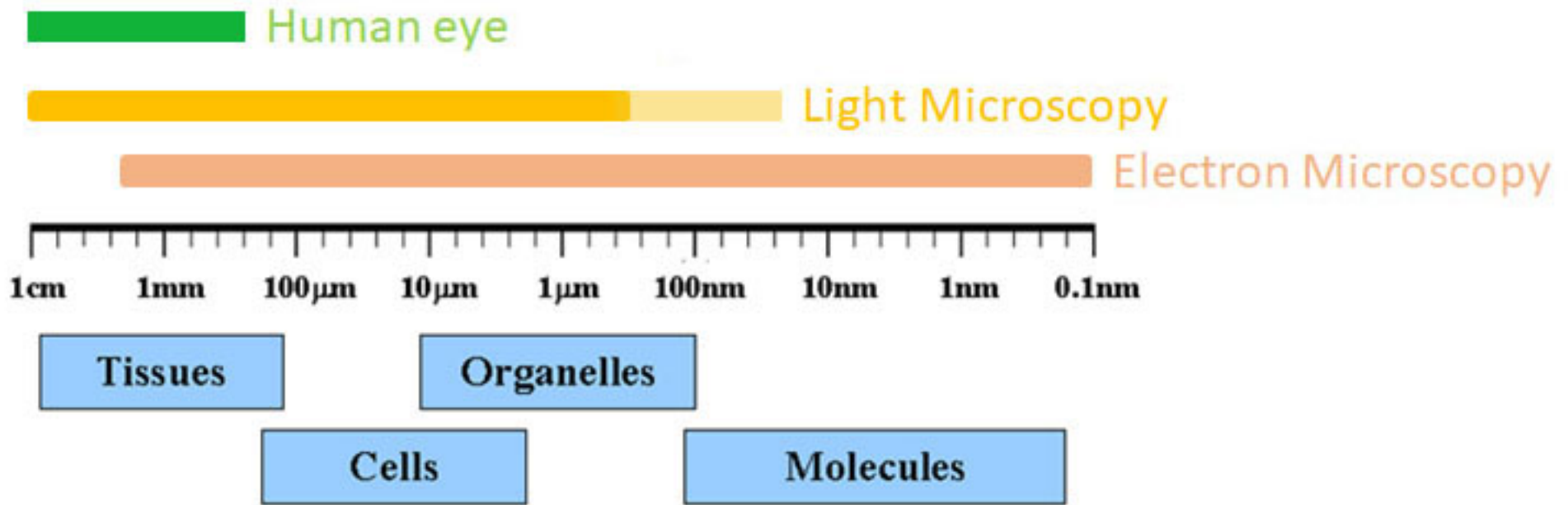
Cell structure of cork by Hooke



Hooke's drawing of a flea



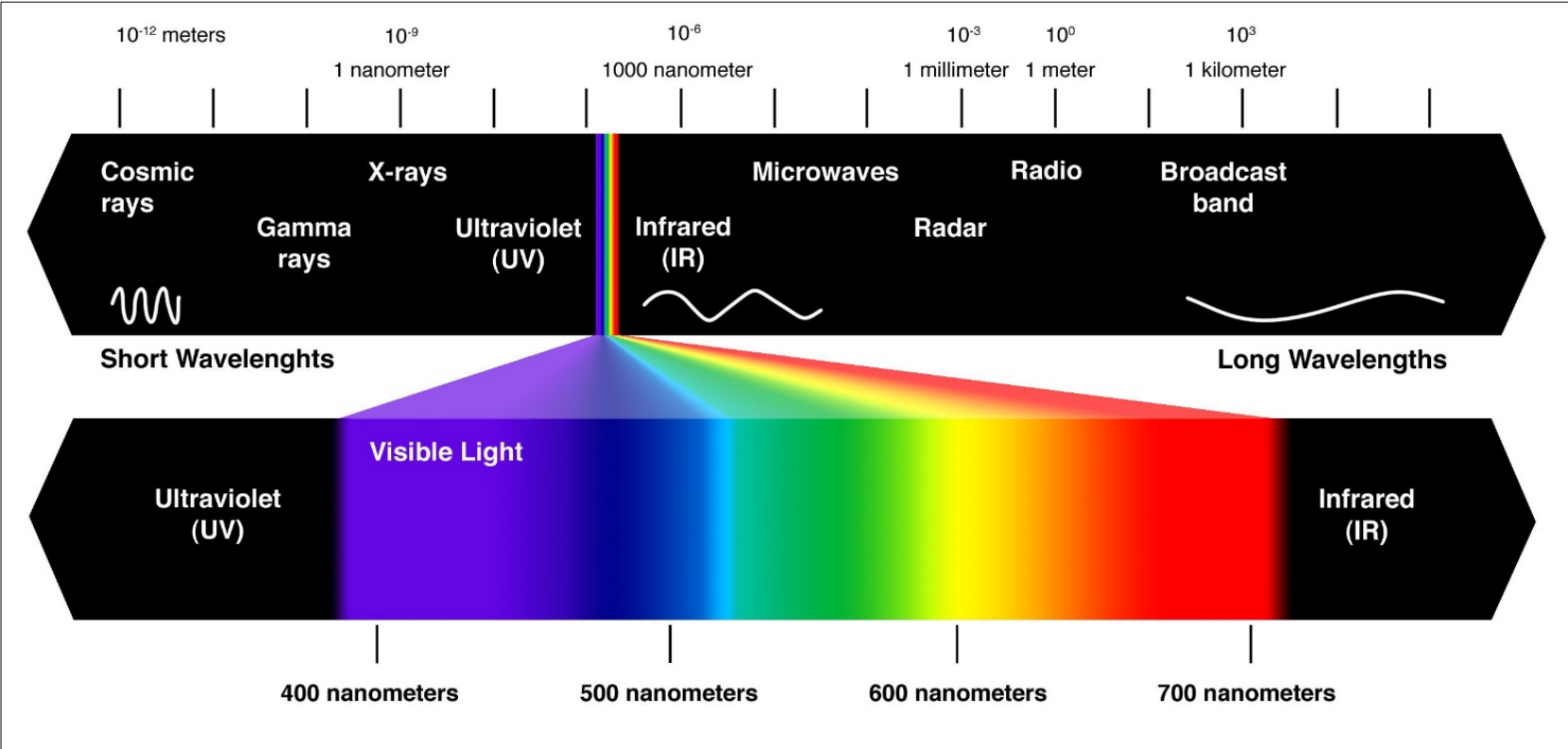
# Bio imaging length scale



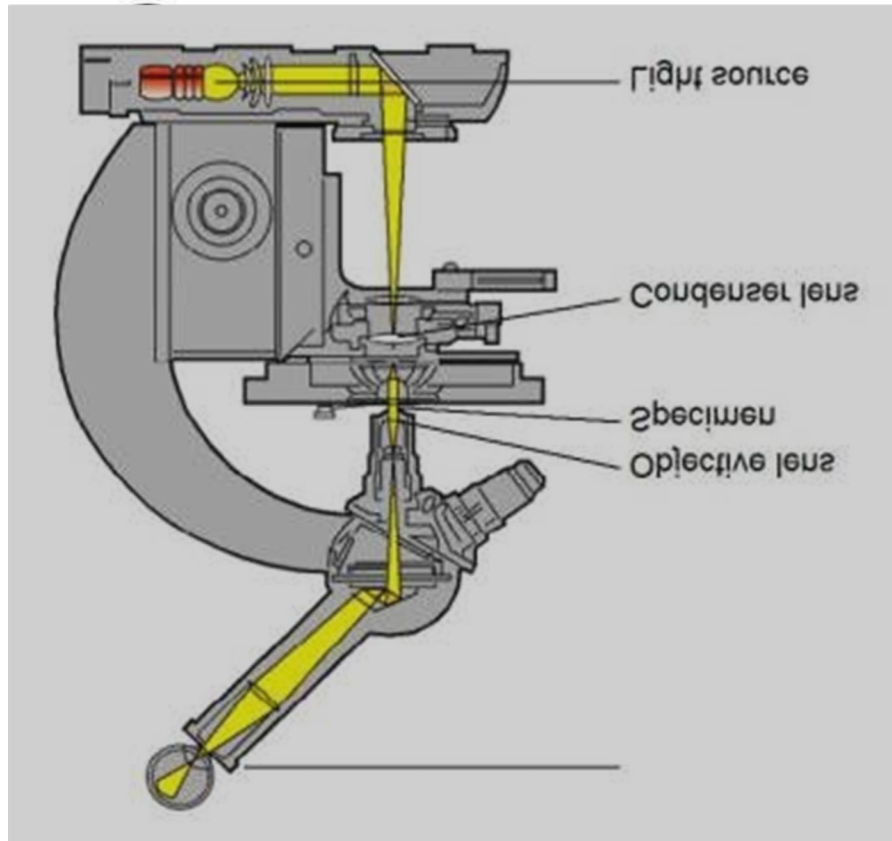
**Light microscopy:**  
Lower resolution  
natural environment  
molecular specificity  
Dynamics  
Wavelength 400-700nm

**Electron microscopy:**  
High resolution  
Vacuum  
Contrast  
Identification  
Wavelength 2-4 nm

# Light Microscopy

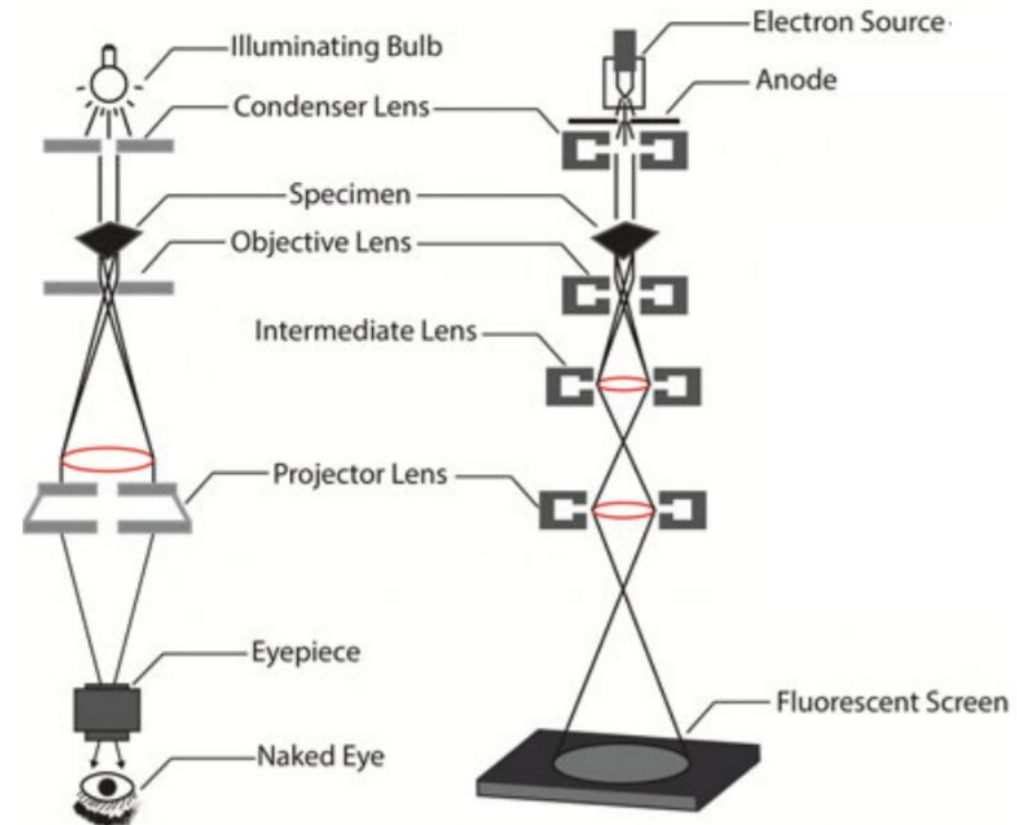


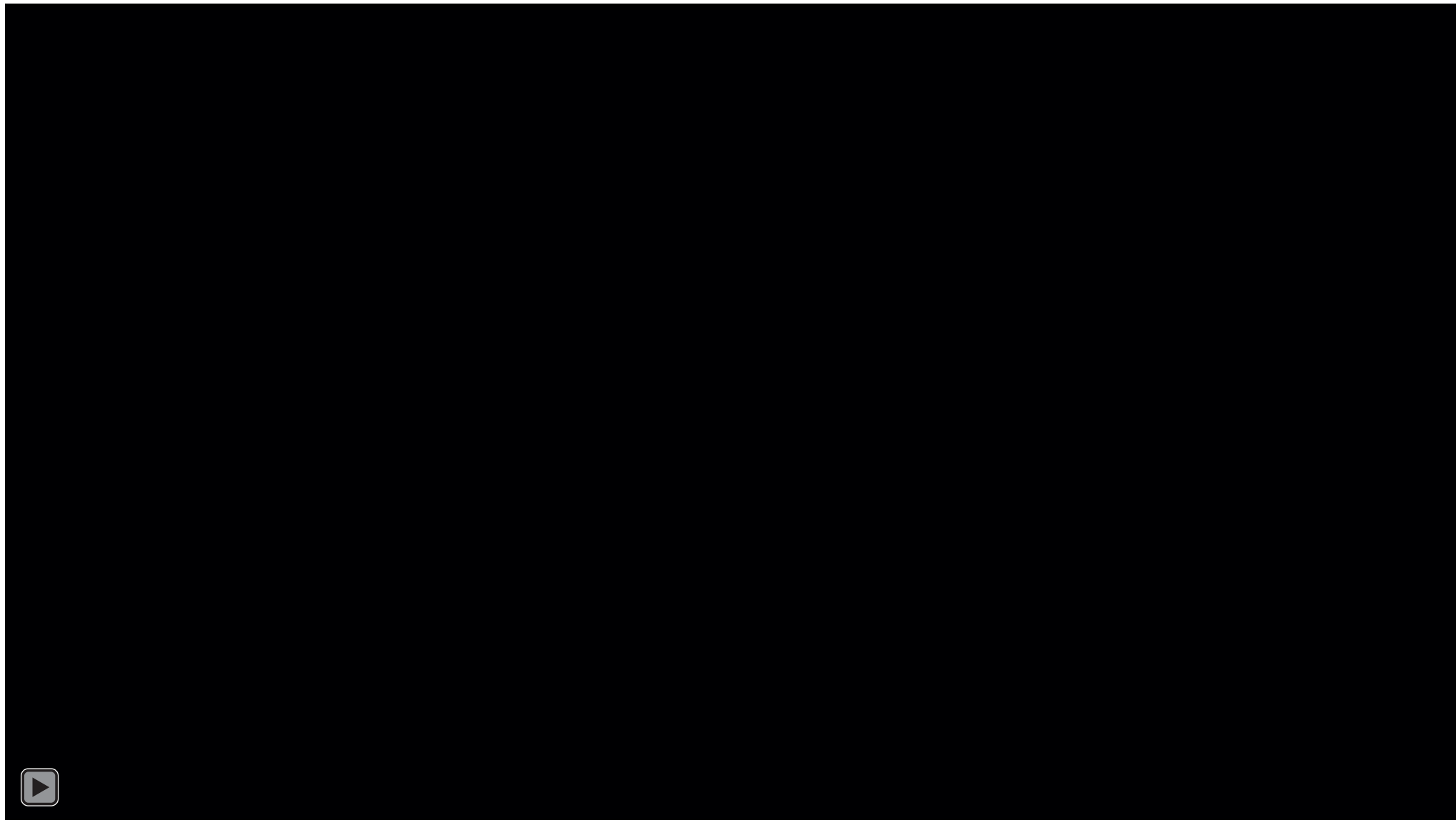
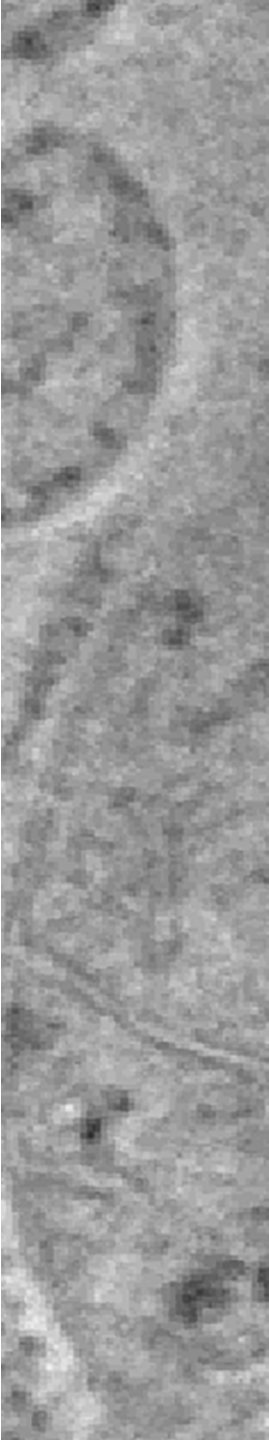
# Compound Microscope



## Light Microscopy

## Transmission Electron Microscopy





# Magnification

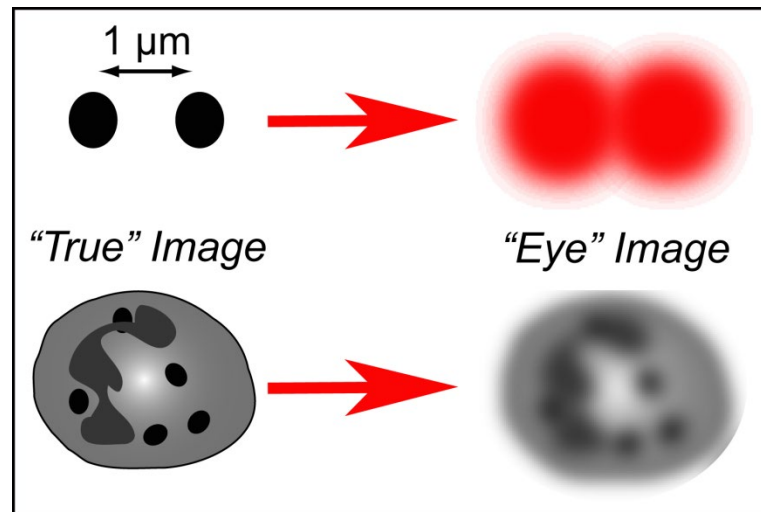
- How high can we go with the magnification?
- Is there a limit?
- What happen to the magnified image?



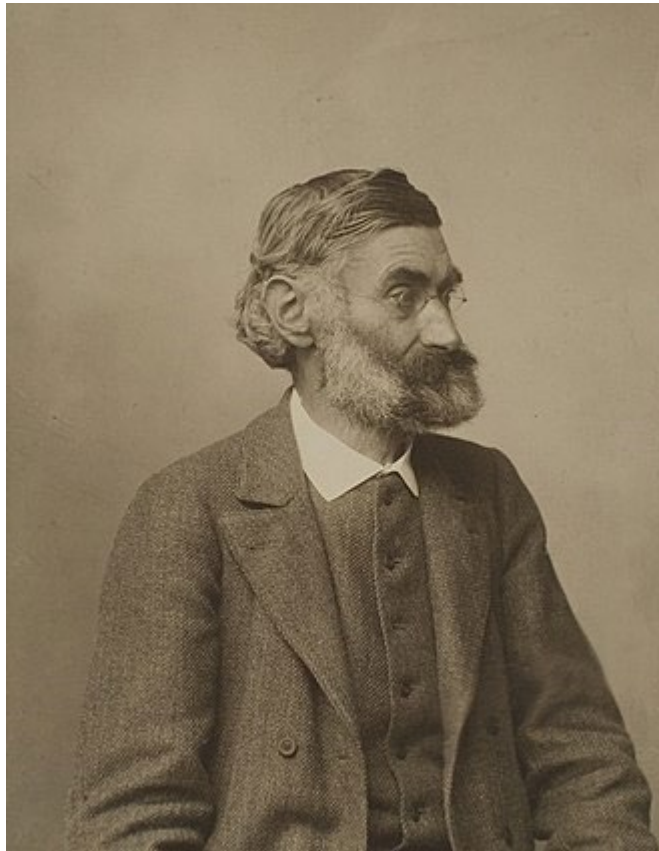


# The Diffraction Limit

- High magnification does not translate into the ability to see infinitely small details. Instead, the resolution of light microscopy is limited because light is a wave and is subject to **diffraction**.
- The **diffraction** of light prevents exact convergence of the rays, causing a sharp point on the object to blur into a **finite-sized spot** in the image.



# Ernst Karl Abbe



Ernst Karl Abbe approximated the diffraction limit of a microscope as 
$$d = \frac{\lambda}{2 n \sin\theta}$$

# Abbe Diffraction Limit

$$d = \frac{\lambda}{2n \sin\theta}$$

NA

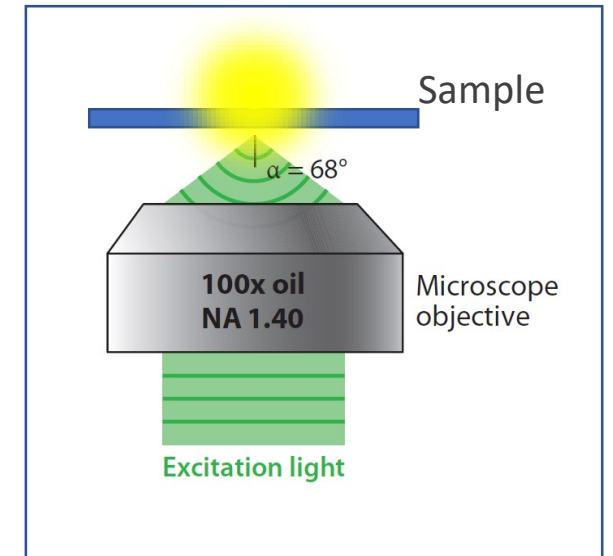
**d** - resolvable feature size

**$\lambda$**  - wavelength

**n** - index of refraction of the medium being imaged in

**$\theta$**  - half-angle of maximum cone of light of the objective

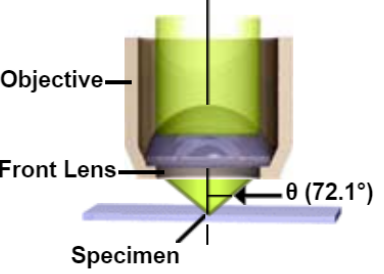
**NA** - objective numerical aperture



In light microscopy the diffraction limit is approximately half the wavelength  $\sim 300\text{nm}$

# Numerical Aperture (NA)

$$NA = n \sin\theta$$



Objective  
Front Lens  
Specimen

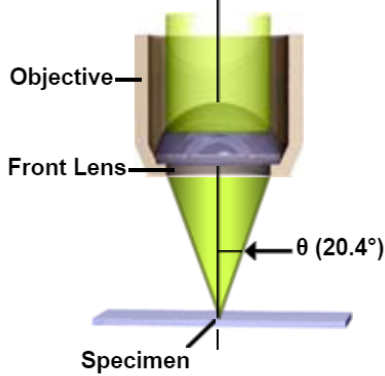
$\theta$  (72.1°)

$NA = n \sin(\theta)$   
 $0.95 = 1.0 \sin 72.1^\circ$

NA = Numerical Aperture  
n = Refractive Index  
= 1.00 (Air)  
 $\theta = 1/2$  Angular Aperture

Approximate Magnification: 100x

Numerical Aperture = 0.95



Objective  
Front Lens  
Specimen

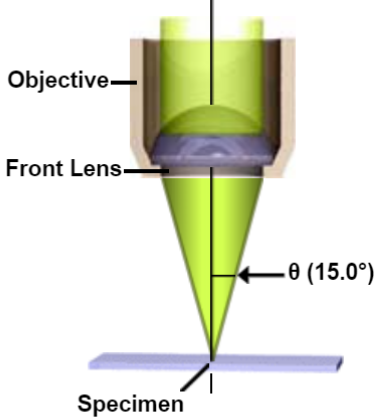
$\theta$  (20.4°)

$NA = n \sin(\theta)$   
 $0.34 = 1.0 \sin 20.4^\circ$

NA = Numerical Aperture  
n = Refractive Index  
= 1.00 (Air)  
 $\theta = 1/2$  Angular Aperture

Approximate Magnification: 20x

Numerical Aperture = 0.34



Objective  
Front Lens  
Specimen

$\theta$  (15.0°)

$NA = n \sin(\theta)$   
 $0.25 = 1.0 \sin 15.0^\circ$

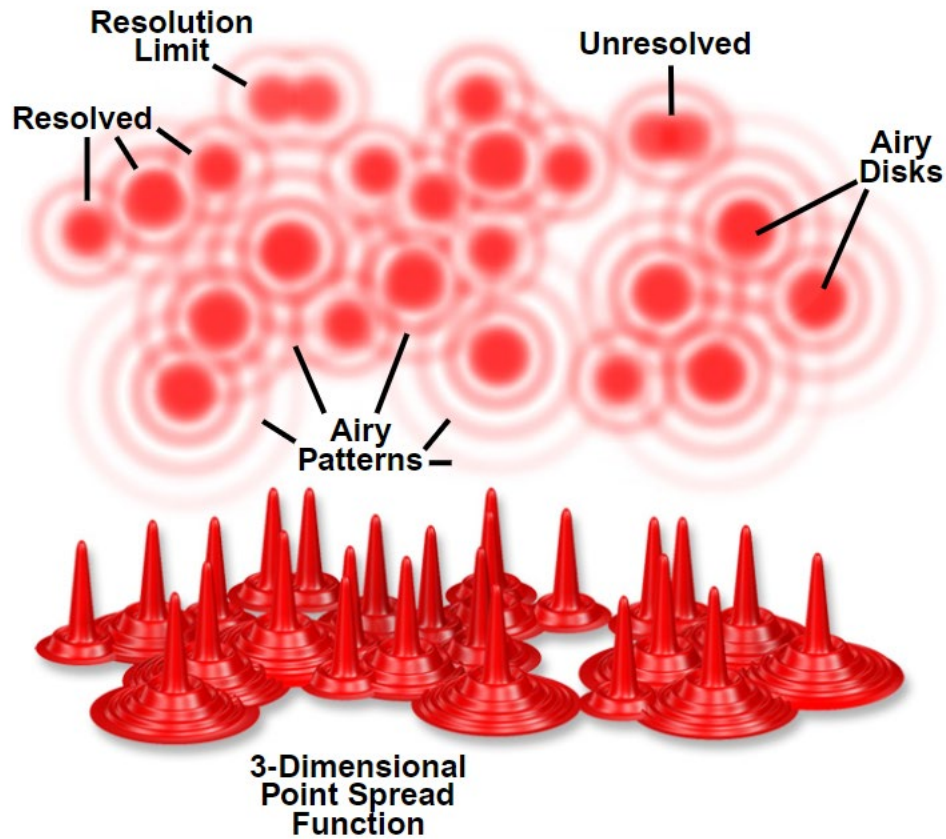
NA = Numerical Aperture  
n = Refractive Index  
= 1.00 (Air)  
 $\theta = 1/2$  Angular Aperture

Approximate Magnification: 10x

Numerical Aperture = 0.25

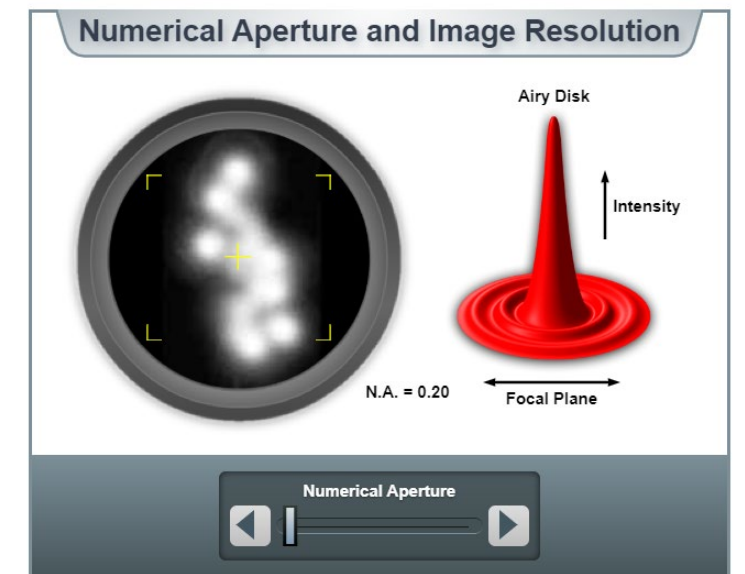
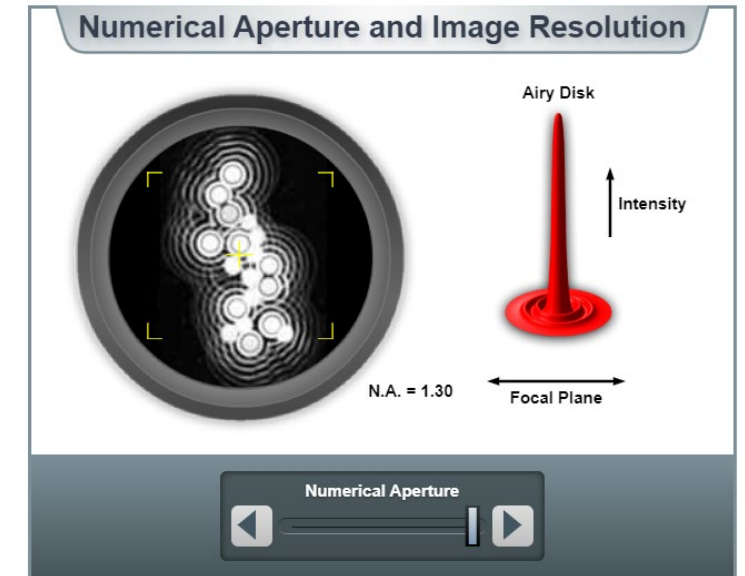
Higher NA  
↓  
Gathering light over larger sets of angles  
↓  
Higher resolution

# Point Spread Function



Three-dimensional representation of the diffraction pattern near the intermediate image plane is known as the **point spread function**

<https://www.microscopyu.com/microscopy-basics/resolution>



# Resolution and not Magnification

## How can we improve the resolution?

- Use shorter wavelength – electron microscopes
- Break the diffraction limit – super resolution microscopy

# Fluorescence Phenomena

- “Fluorescence”: named by George Gabriel Stokes (1852) after the mineral fluorite which lights up when illuminated with UV.
- He realized that the exciting light wavelength will always be shorter than the emitted light wavelength.
- The Stokes shift, which describes this light conversion, is named in Stokes's honor.



Fluorite



George Gabriel Stokes (1818-1903)

# What is Fluorescence?



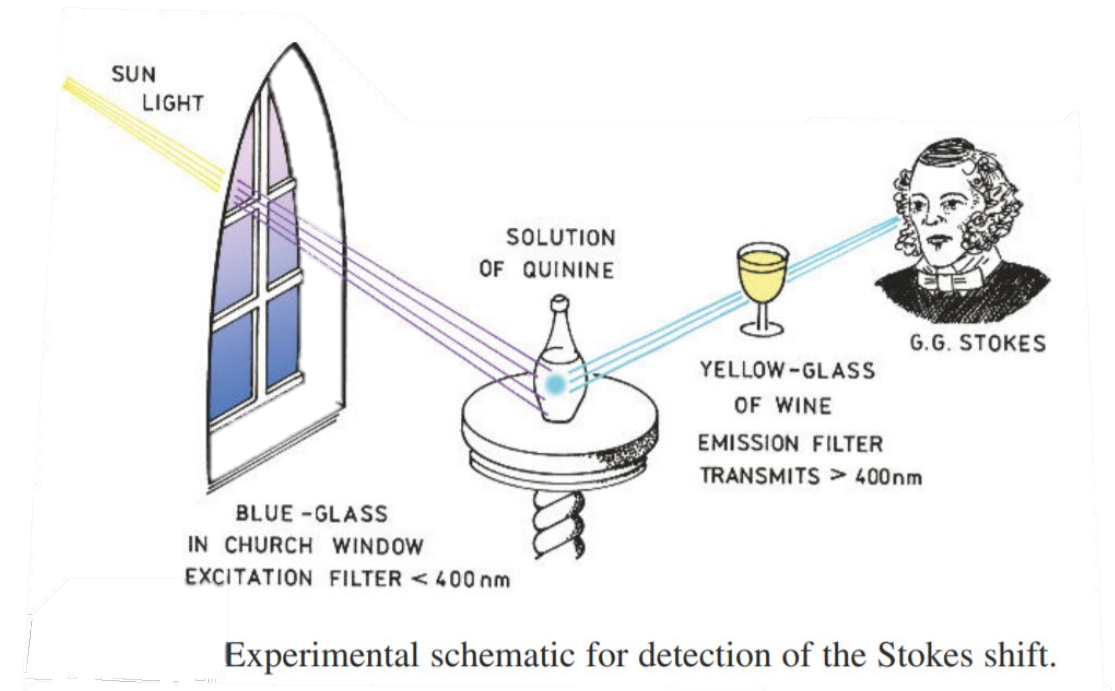
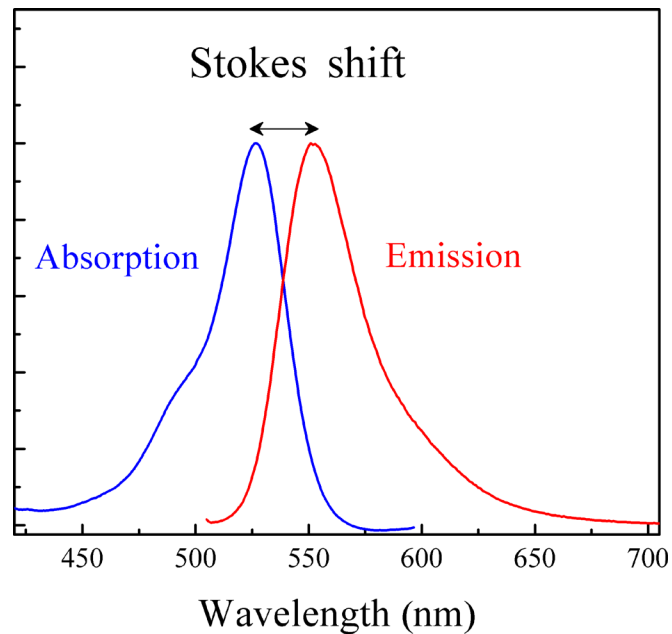
Tonic water (quinine)

Shining light on some molecules, excitation light, results in light emission at a longer wavelength.

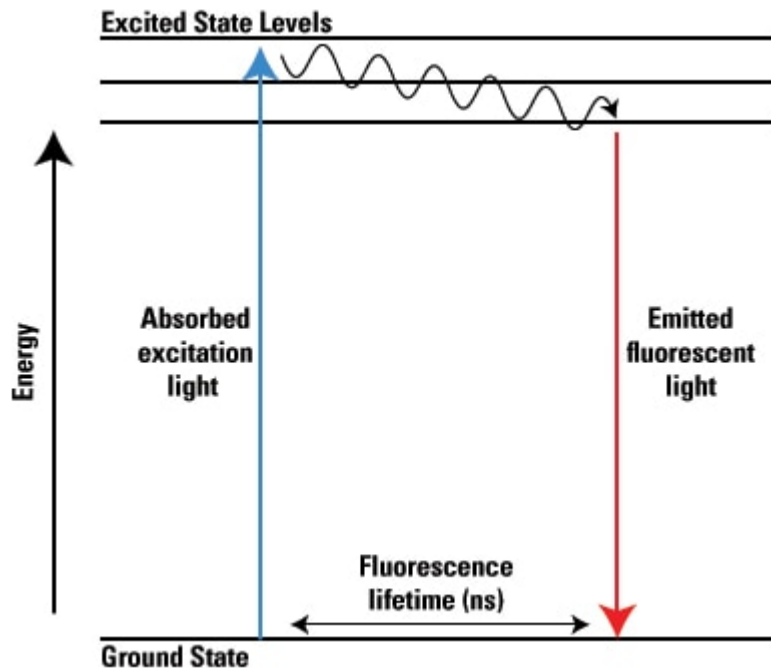


# Stokes Shift

The energy of the emission is typically less than that of absorption. Fluorescence typically occurs at lower energies or longer wavelengths.



# Jablonski Diagram

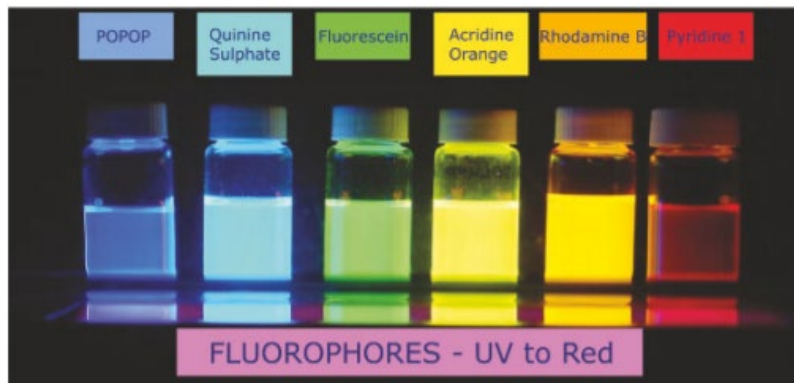
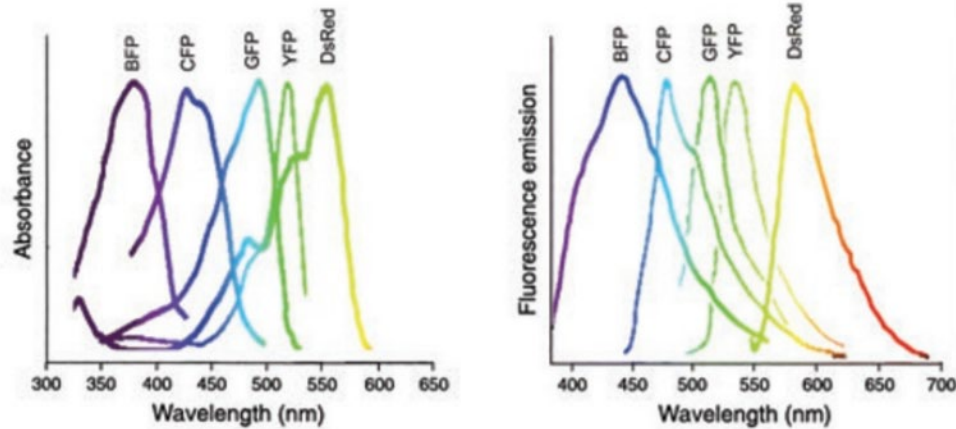


## Fluorescein

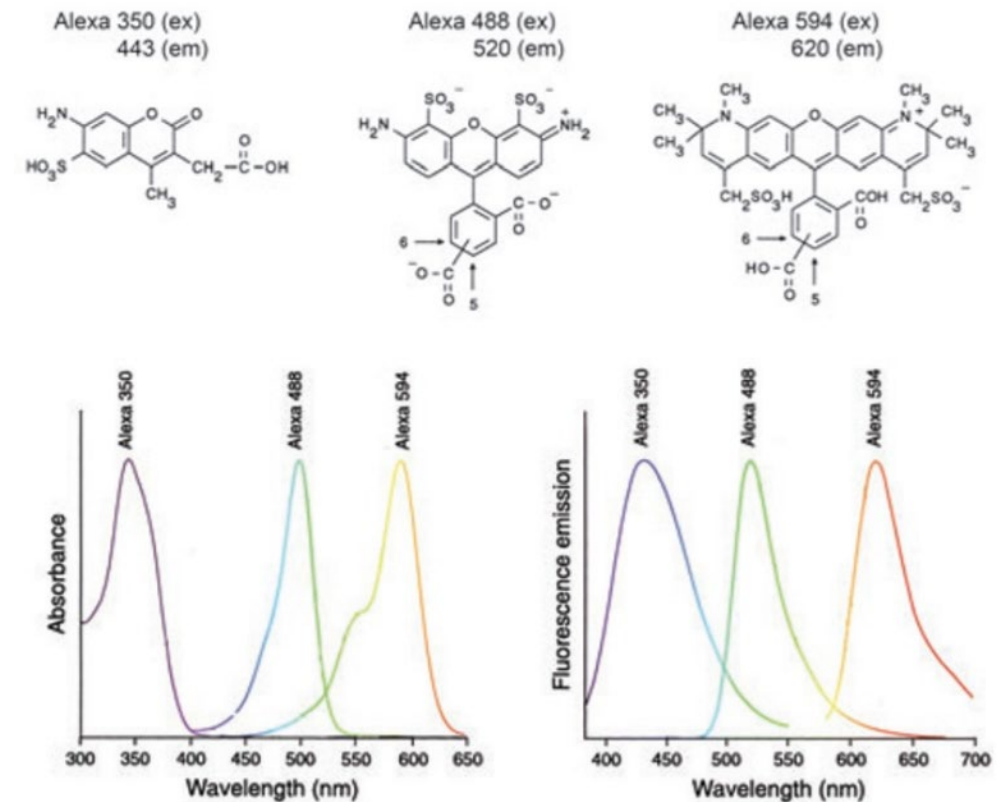


Shining light on some molecules, excitation light, results in light emission at a longer wavelength.

## Fluorescent proteins



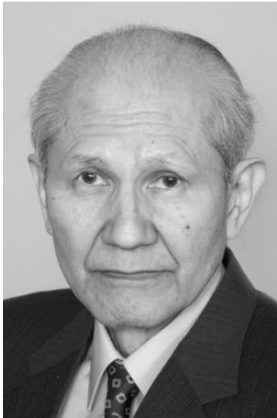
## Fluorescent organic molecules



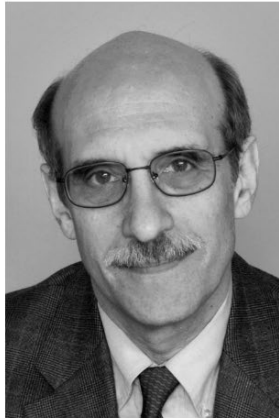
Structural basis of fluorophores is conjugated double bonds acting as 'antenna'

# Green Fluorescent Protein (GFP)

## The Nobel Prize in Chemistry 2008



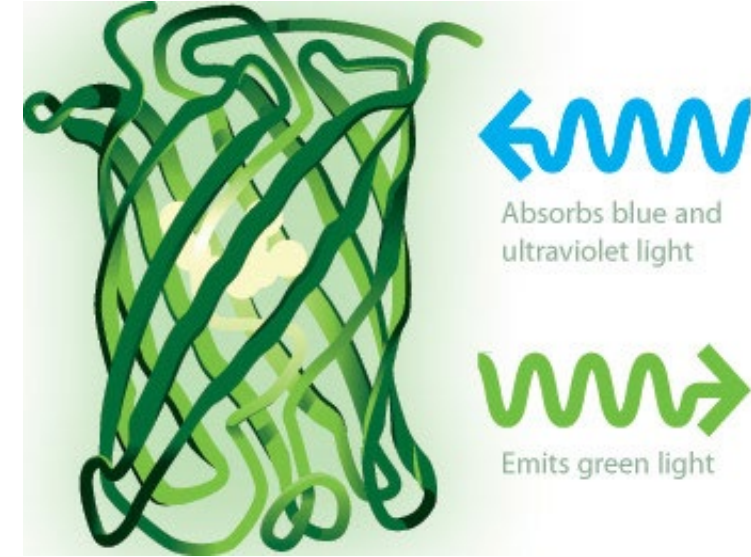
© The Nobel Foundation. Photo: U. Montan  
Osamu Shimomura  
Prize share: 1/3



© The Nobel Foundation. Photo: U. Montan  
Martin Chalfie  
Prize share: 1/3



© The Nobel Foundation. Photo: U. Montan  
Roger Y. Tsien  
Prize share: 1/3



**Osamu Shimomura** first isolated GFP from the jellyfish *Aequorea victoria*, and discovered that it glowed bright green under ultraviolet light.

**Martin Chalfie** demonstrated the value of GFP genetic tag for biological phenomena

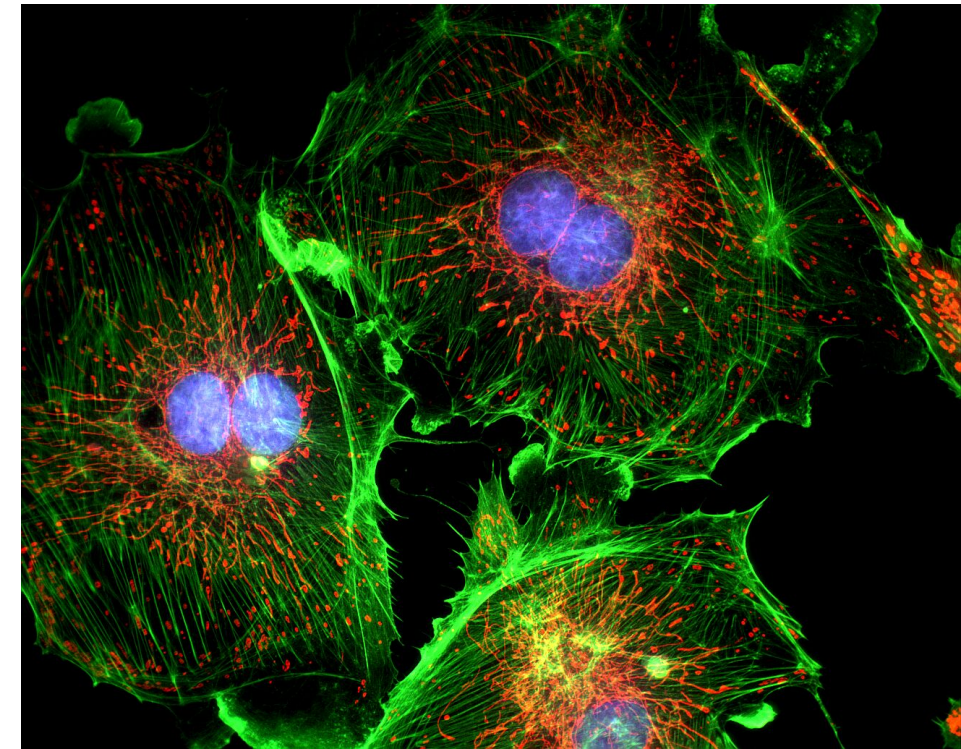
**Roger Y. Tsien** contributed to our general understanding of how GFP fluoresces and extended the color palette

“For the discovery and development of the green fluorescent protein, GFP.”

# Fluorescence Microscopy

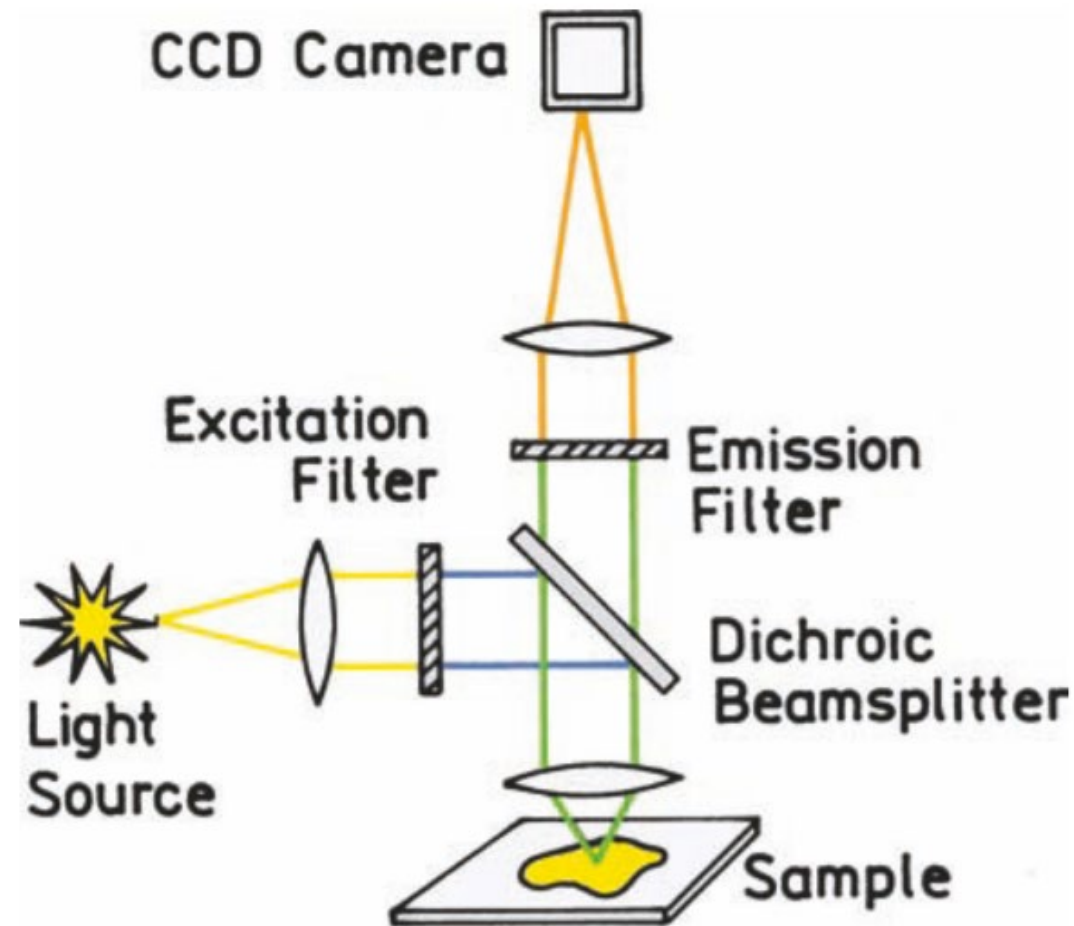
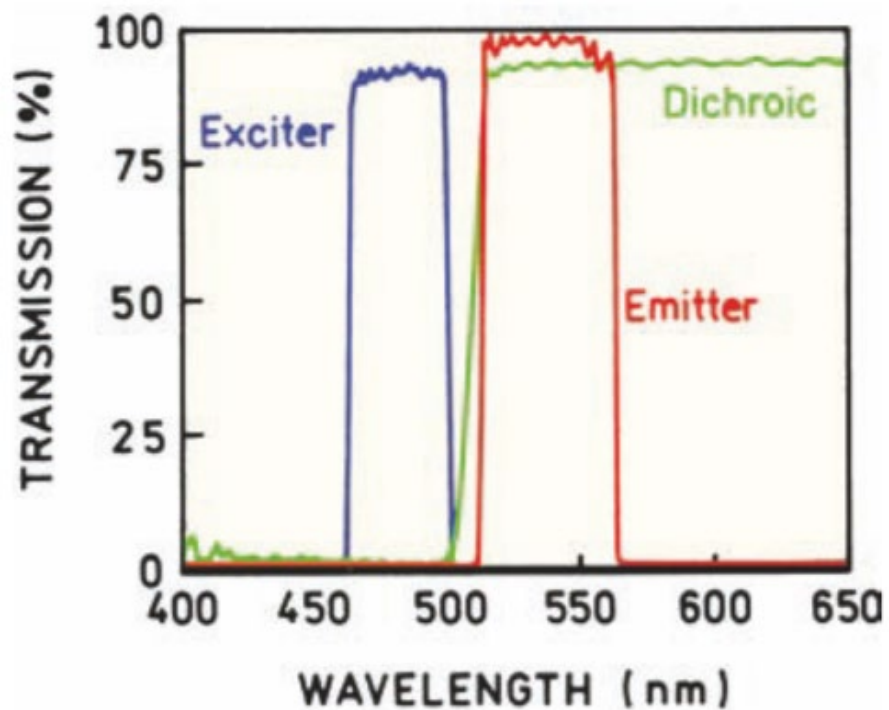
## Why Fluorescence?

- High specificity
  - Fluorescent proteins (GFP...)
  - Antibodies conjugated to fluorescent molecules
- High contrast
  - Bright signal on dark background
- Quantitative
- Live cell imaging – dynamics
- Natural imaging conditions



# Filters

Epifluorescence filter set for Cy5



# Super Resolution Microscopy



The Nobel Prize in Chemistry 2014  
Eric Betzig, Stefan W. Hell, William E. Moerner

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## The Nobel Prize in Chemistry 2014



Photo: Matt Staley/HHMI

**Eric Betzig**

Prize share: 1/3



© Bernd Schuller, Max-Planck-Institut

**Stefan W. Hell**

Prize share: 1/3



Photo: K. Lowder via Wikimedia Commons, CC-BY-SA-3.0

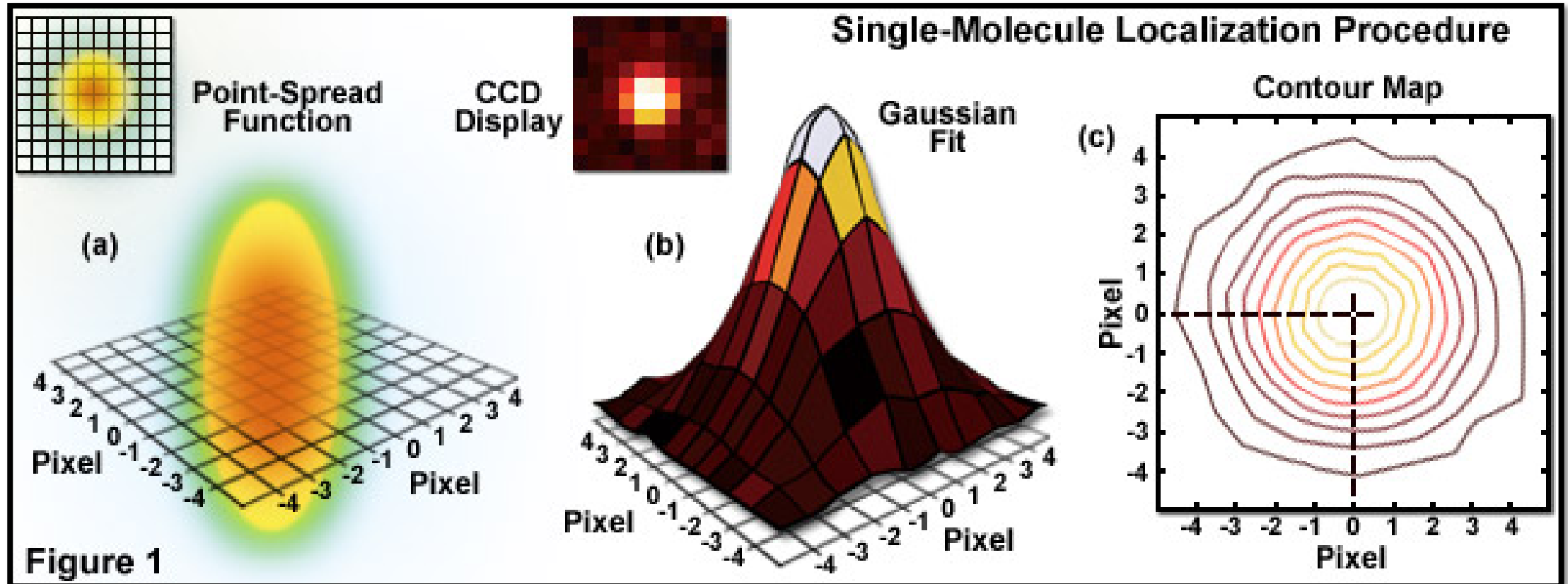
**William E. Moerner**

Prize share: 1/3

The Nobel Prize in Chemistry 2014 was awarded jointly to Eric Betzig, Stefan W. Hell and William E. Moerner *"for the development of super-resolved fluorescence microscopy"*.

# Stochastic Optical Reconstruction Microscopy (STORM)

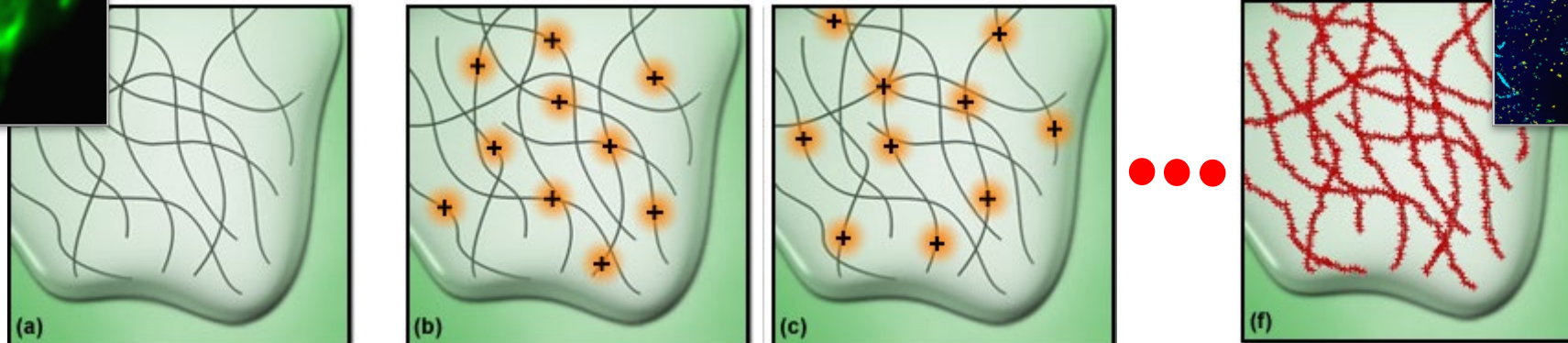
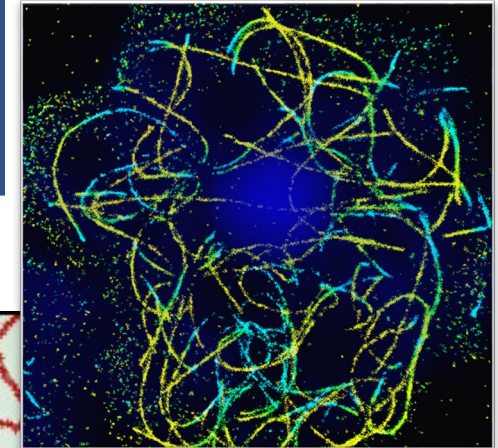
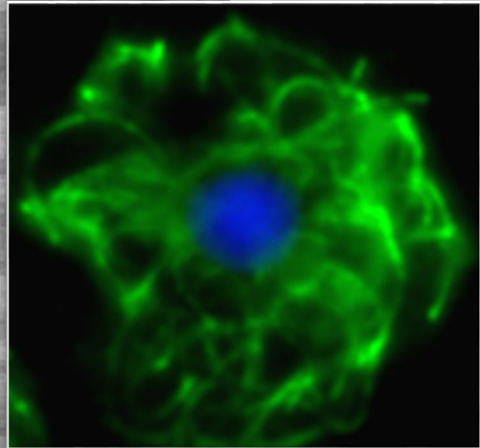
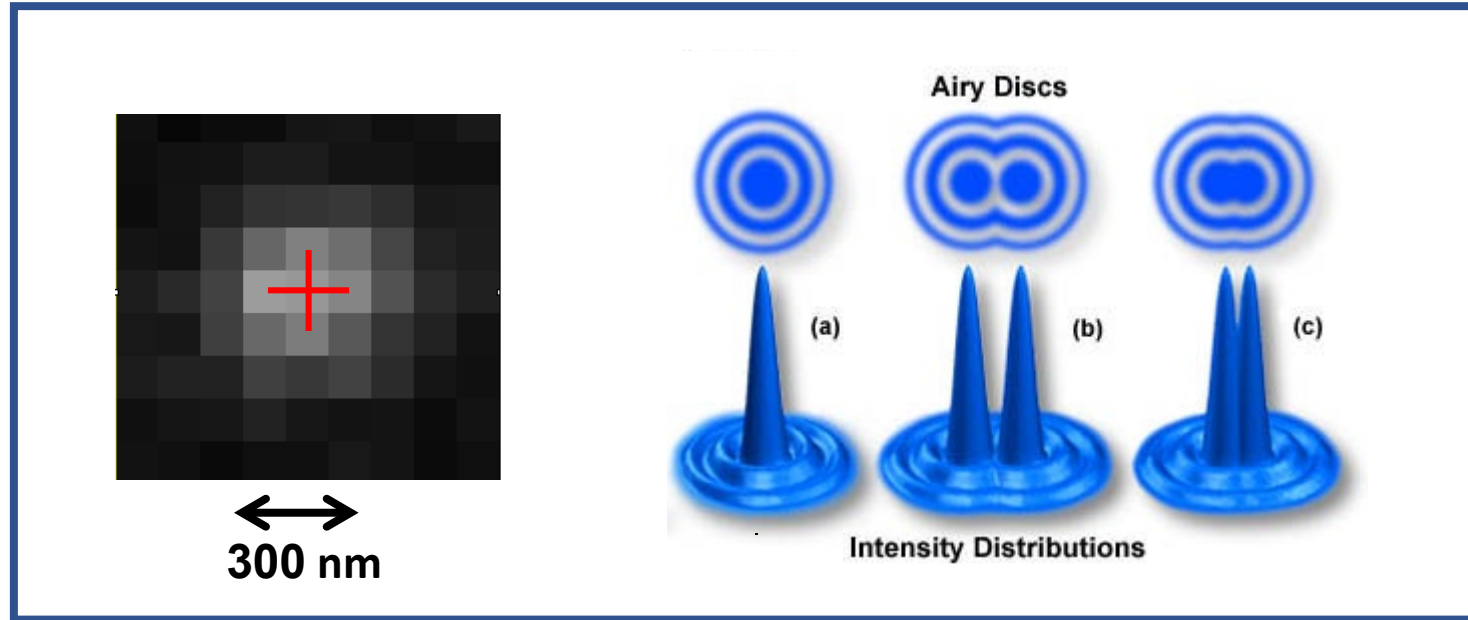
## Point Spread Function (PSF)



- PSF describes the response of an imaging system to a point source.
- Fitting the image using Gaussian function allows to determine the center of the spot with about an order of magnitude higher resolution.



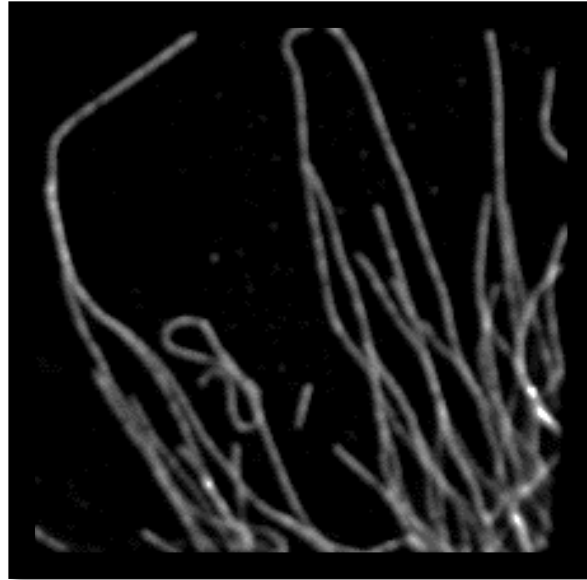
# Stochastic Optical Reconstruction Microscopy (STORM)



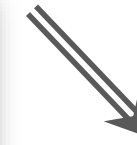
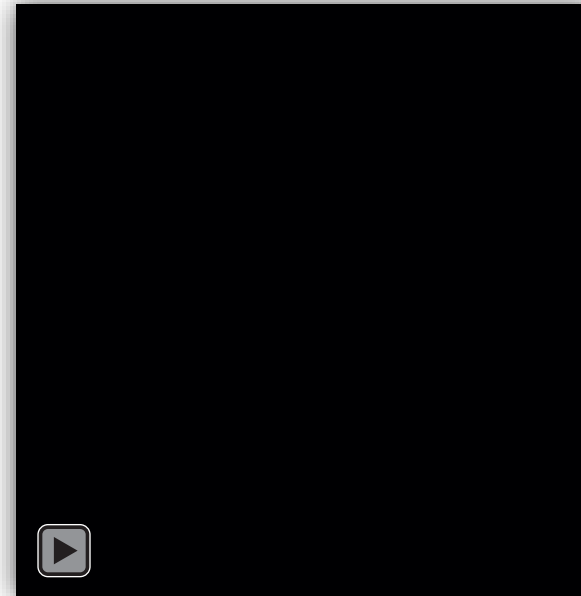
Separation of the molecules is done by switching them ON and OFF stochastically.

# Stochastic Optical Reconstruction Microscopy (STORM)

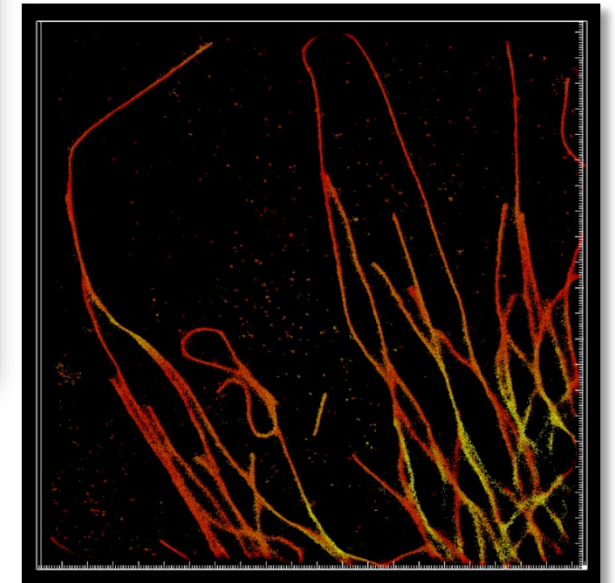
## Conventional Fluorescence



## Blinking



## Super Resolution

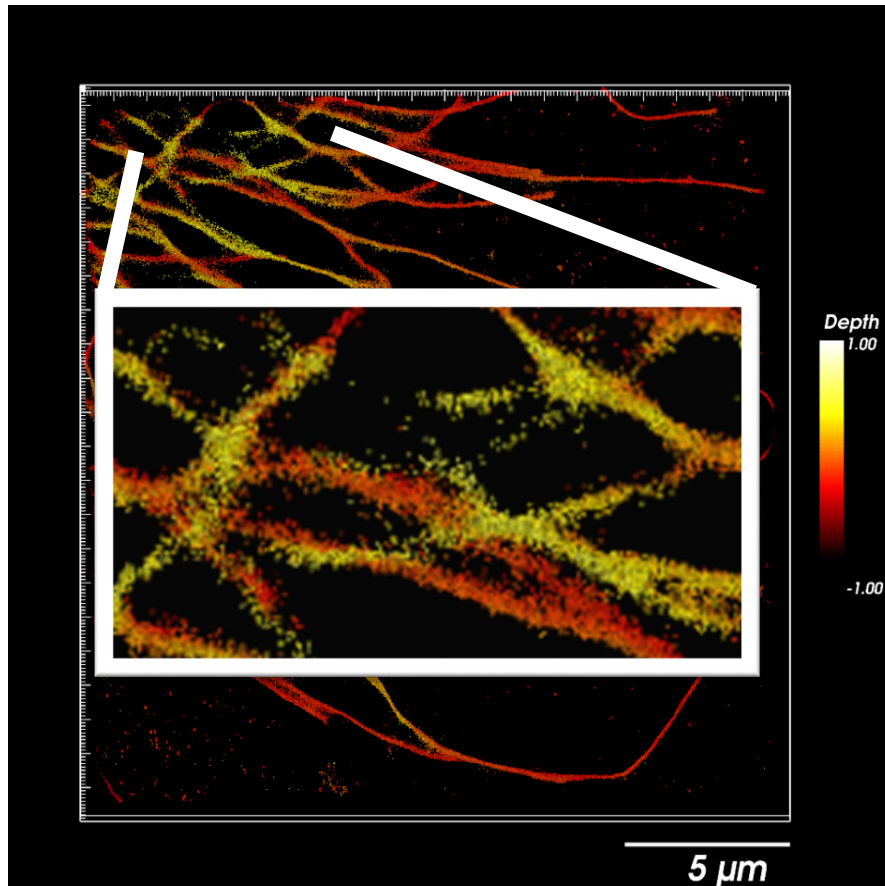


### Vutara 352 (Bruker)

- Lateral resolution of 20 nanometers
- Axial resolution of 50 nanometers
- Up to 5-micron imaging depth (with z-stack acquisition)
- Simultaneous 2-color imaging in super-resolution mode (up to 4 colors in wide-field mode)
- 3D particle tracking with  $\sim 10\text{nm}$  precision

# 3D STORM - microtubules

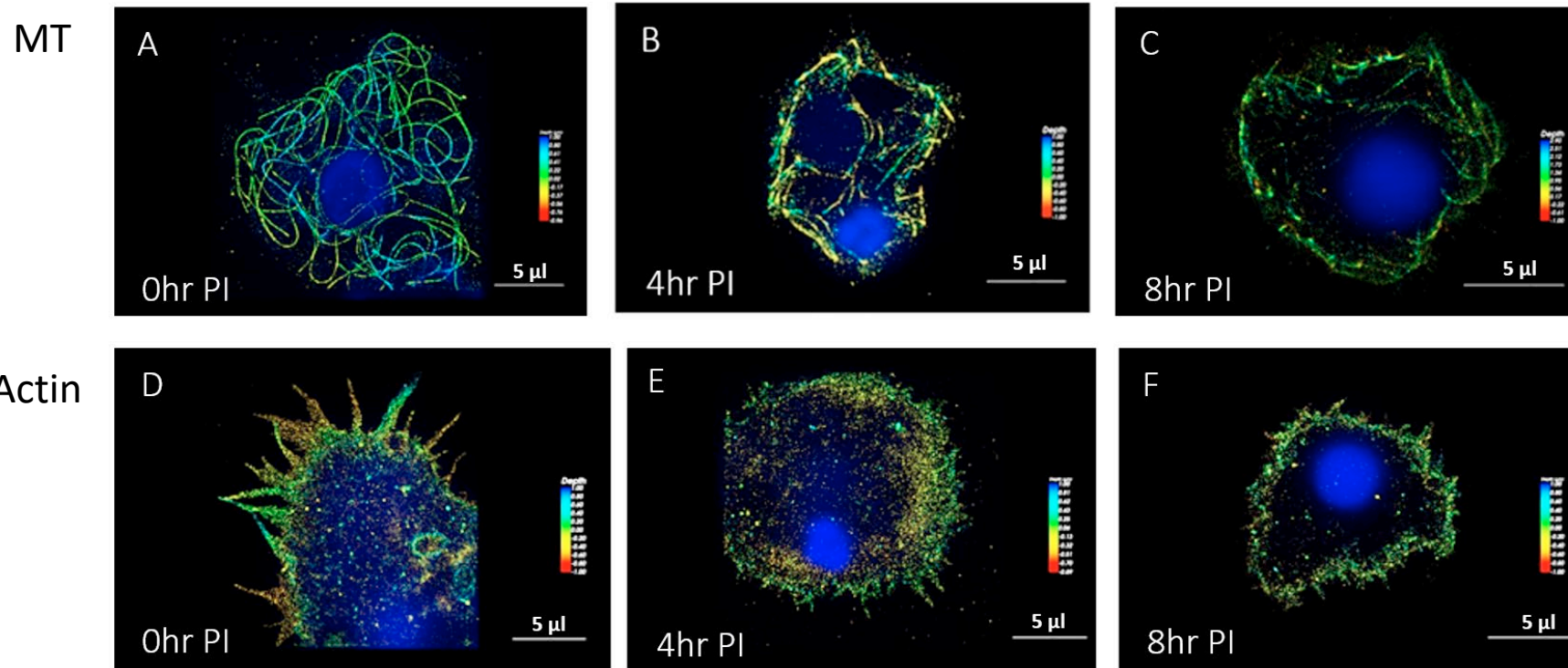
Super resolution



Conventional fluorescence

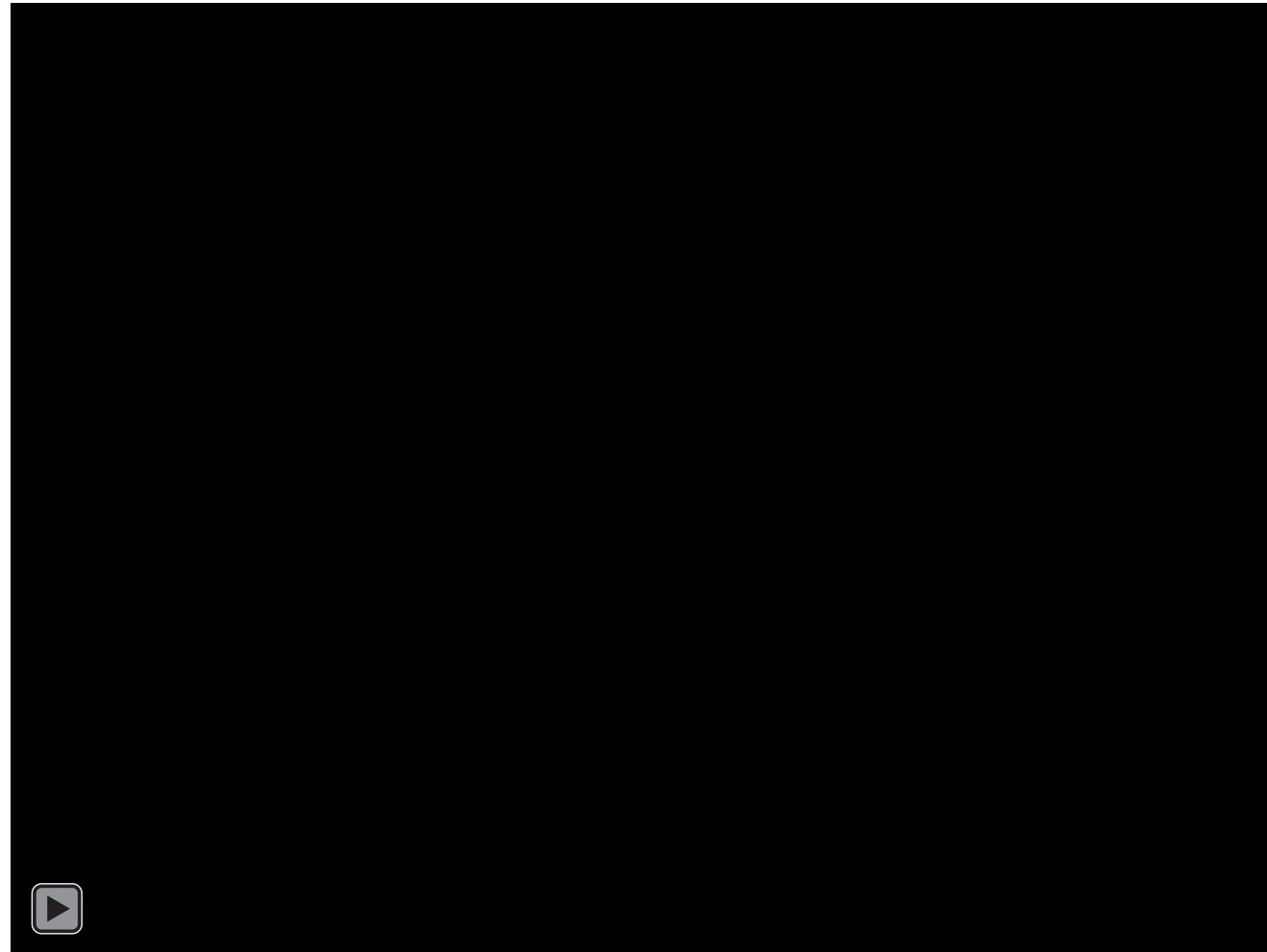
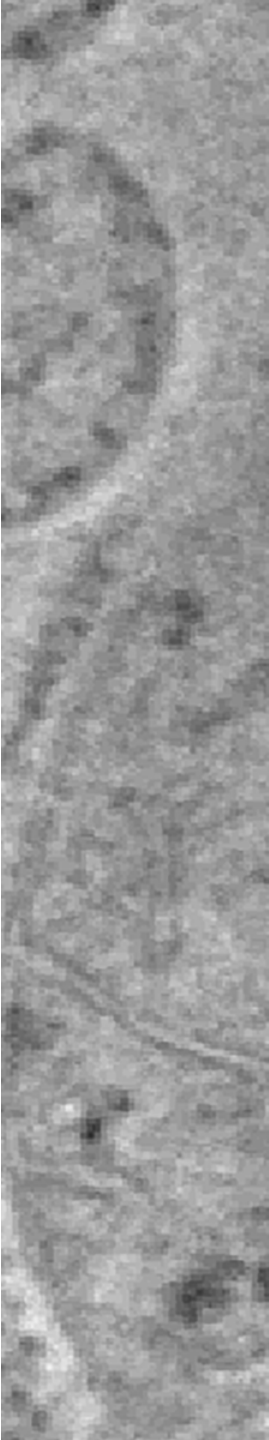


# Morphological changes of the cytoskeleton in Mimivirus-infected cells



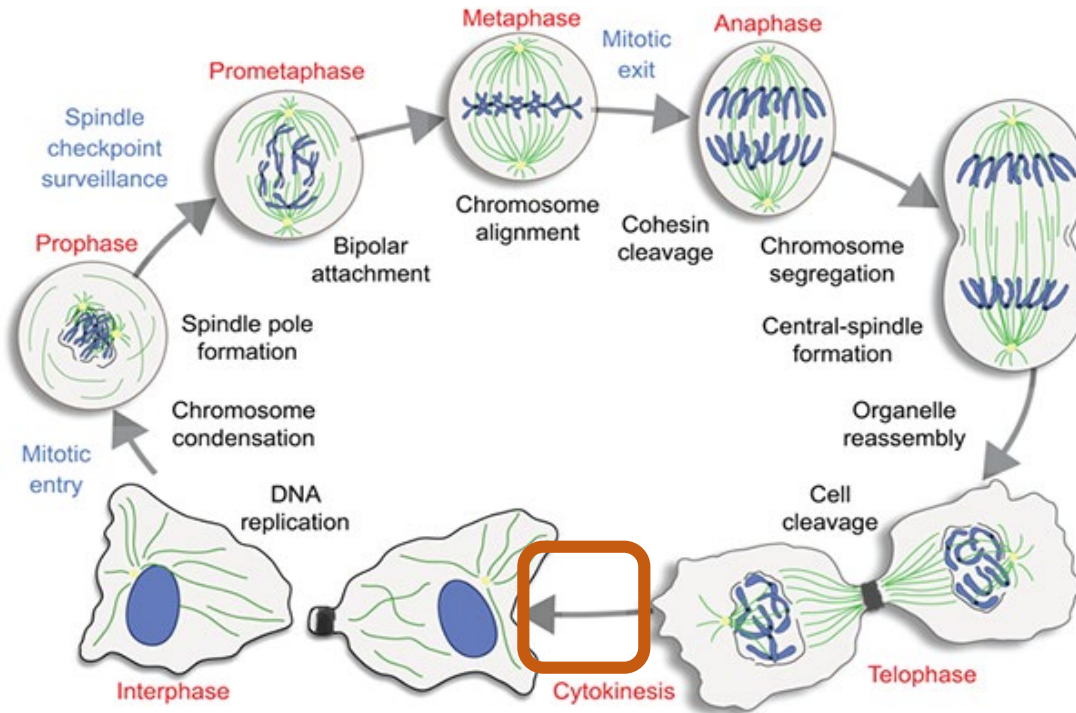
Actin and microtubule filaments rearrangements throughout the infection cycle of *Acanthamoeba* at different time points post infection.

The cells were infected, followed by fixation and staining of microtubules using anti-alpha-tubulin antibodies (A,B,C) or staining of actin fibers using Phalloinin-647 (D,E,F), and DNA using DAPI (blue).

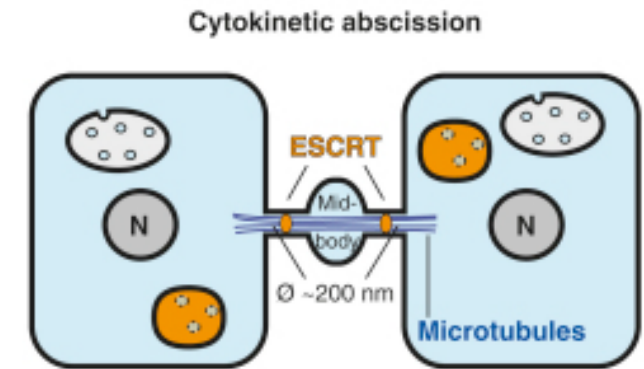


## Cytokinetic abscission

**ESCRT** (Endosomal Sorting Complex Required for Transport) proteins play a role in biogenesis of multi vesicular bodies, HIV budding and cytokinesis.

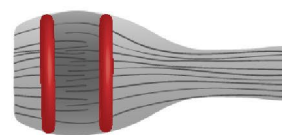
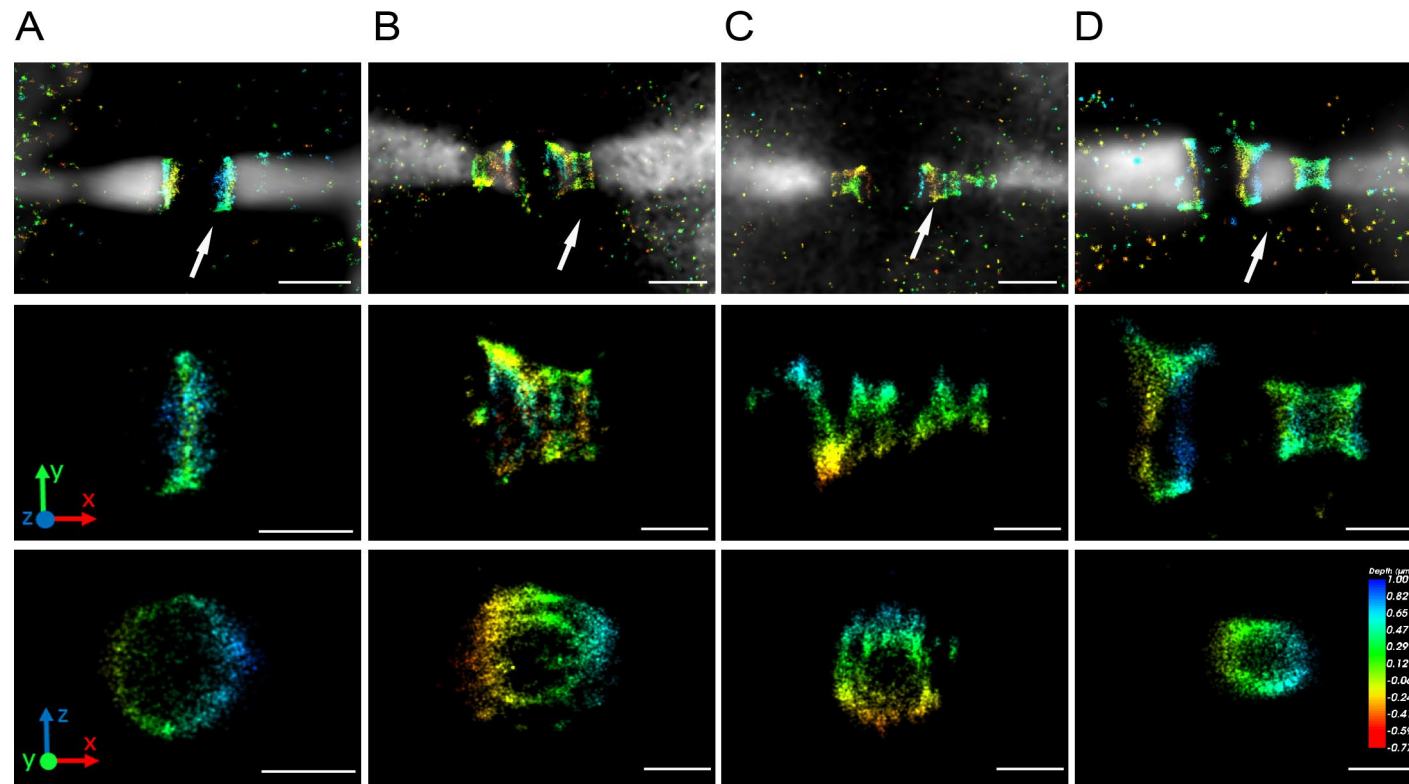


**ESCRT** machinery involves in the final scission of the bridge connecting the two daughter cells

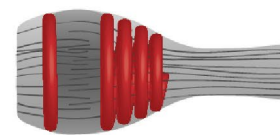


Oliver Schmidth, current Biology 2011

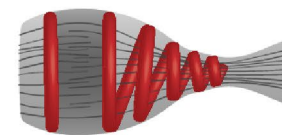
# Spatial organization of the ESCRT-III protein (IST1) in the intercellular bridge of dividing cells at different stages of abscission



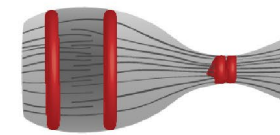
ring assembly



spiral formation

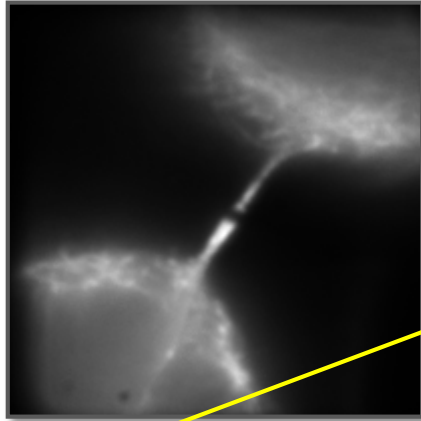


spiral rearrangement

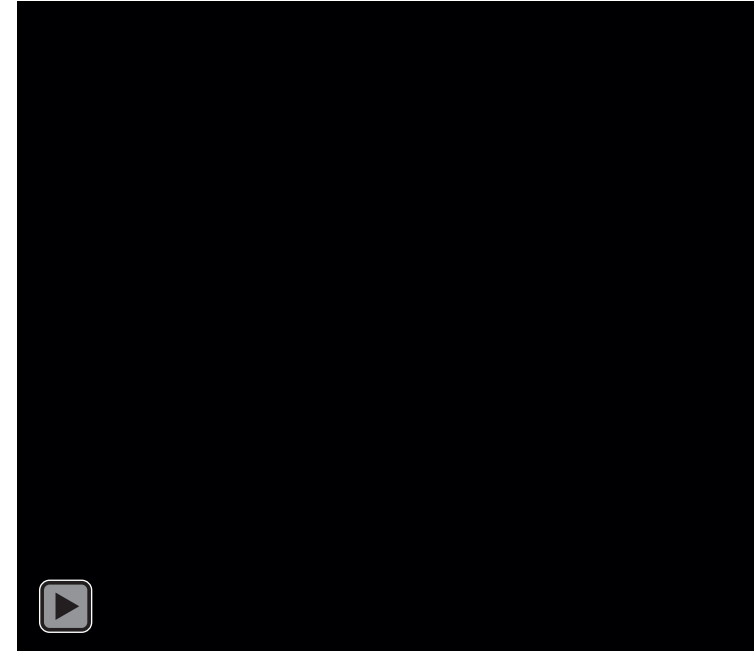
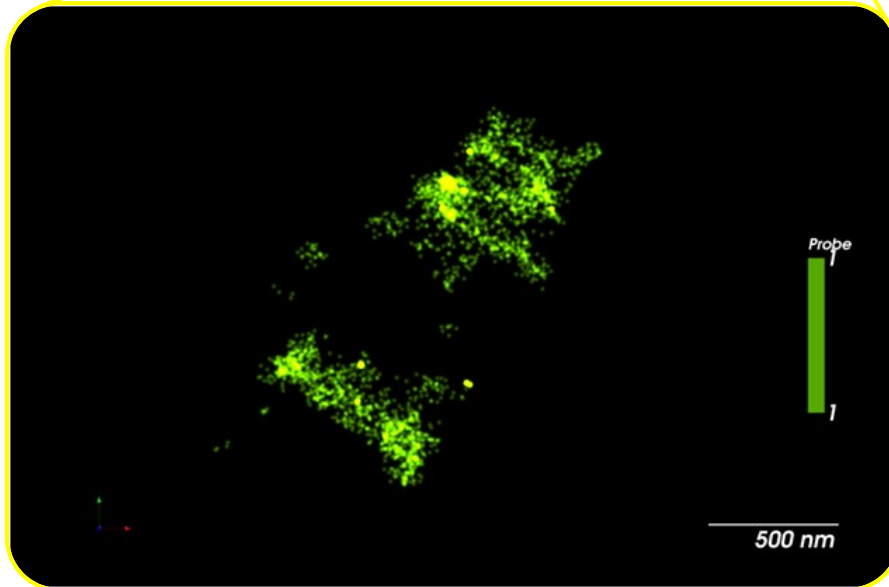
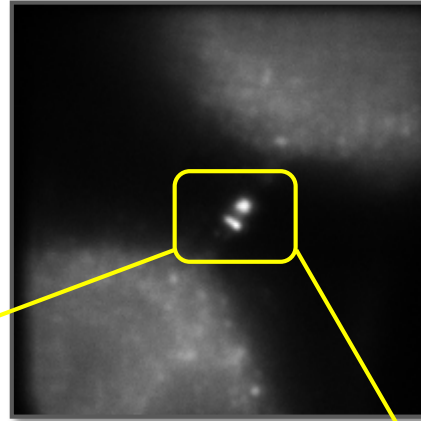


spiral dissociation

Tubulin Alexa488



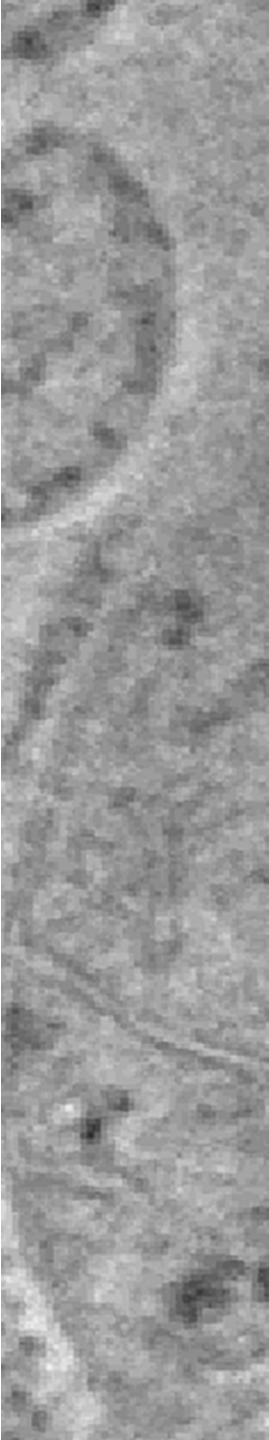
IST1 Alexa647



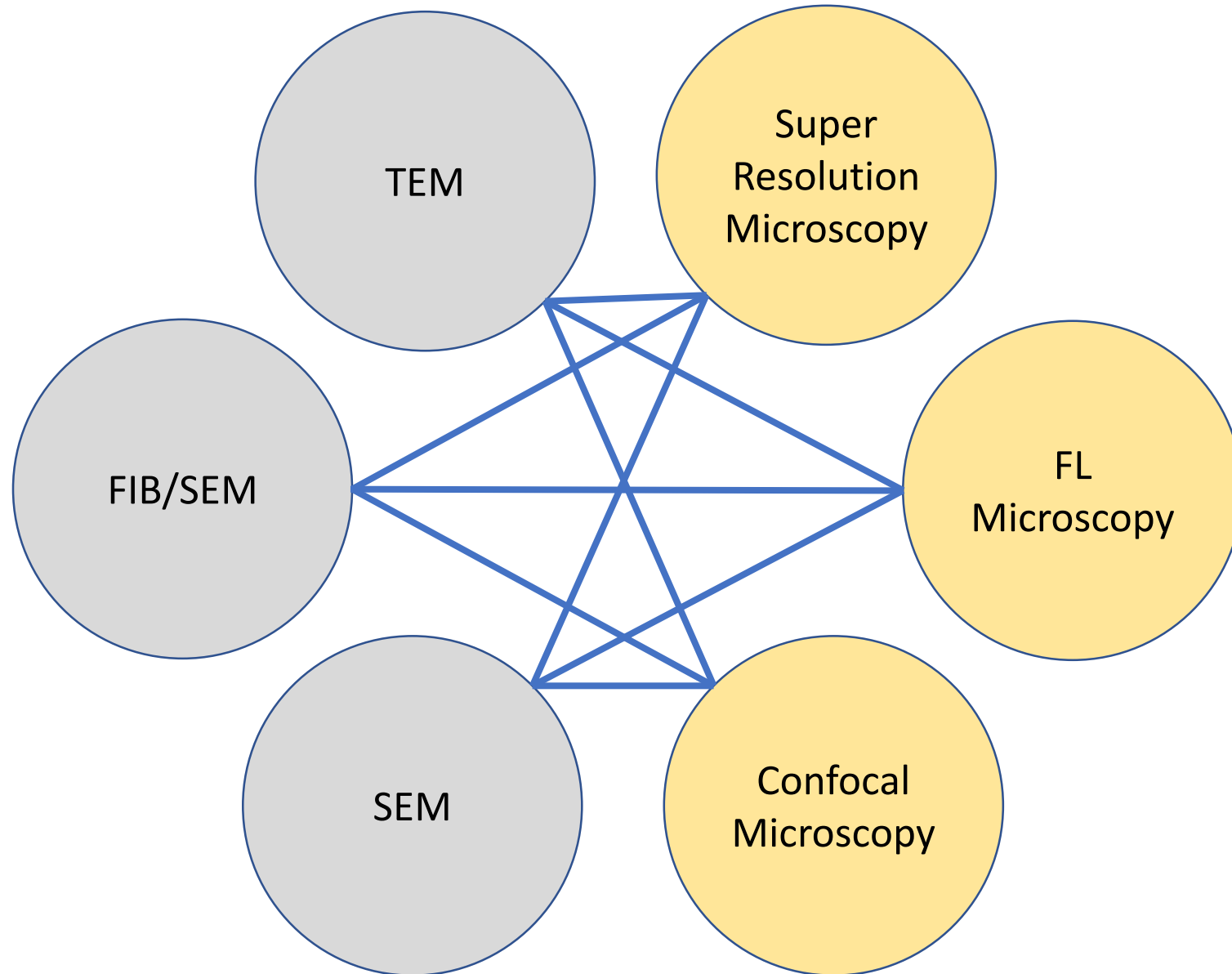
The first direct evidence that the ESCRT –III create helical filaments!!!

*Dr. Natalie Elia and Inna Goliand, BGU*

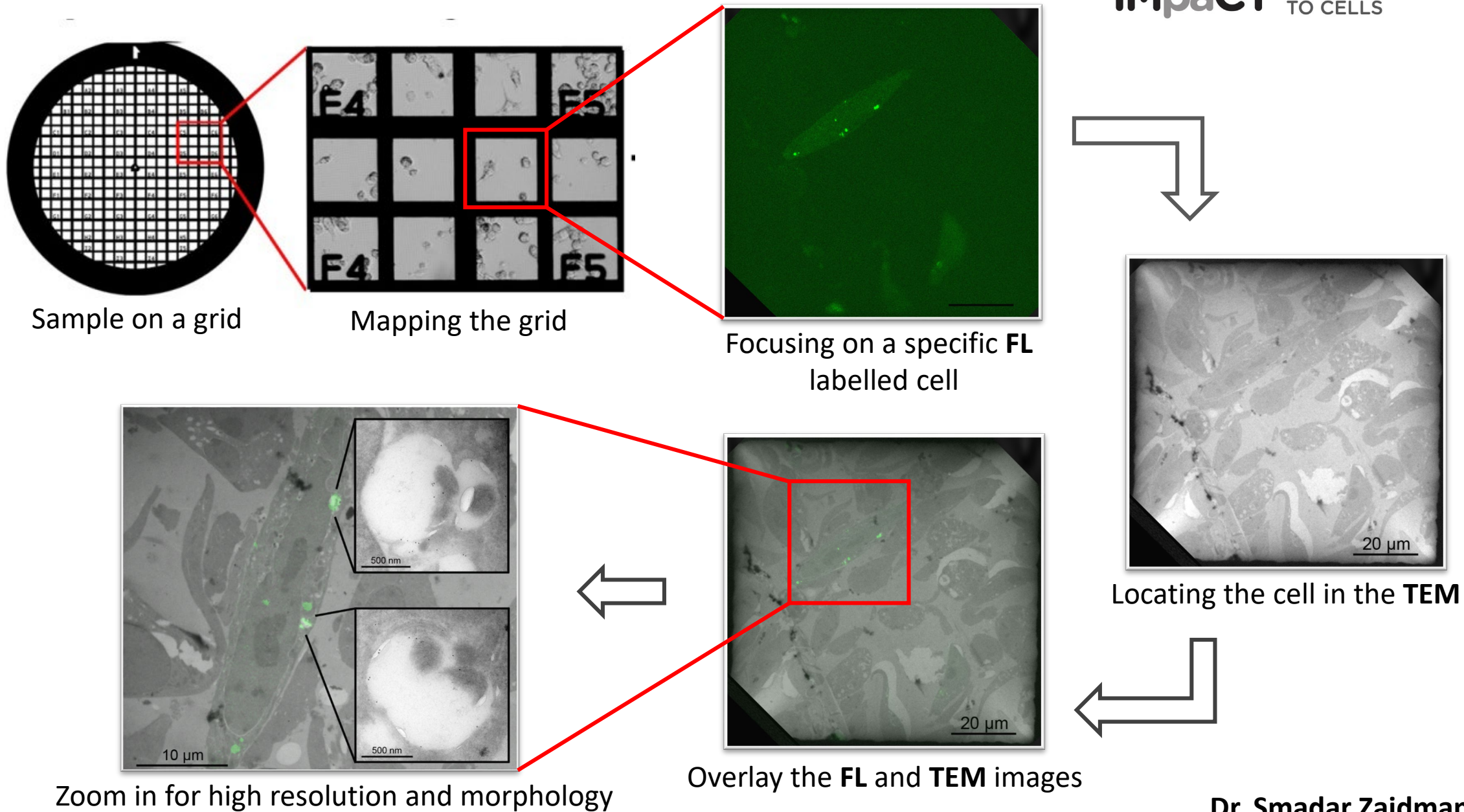




# Correlative Light and Electron Microscopy (CLEM)



# General CLEM workflow



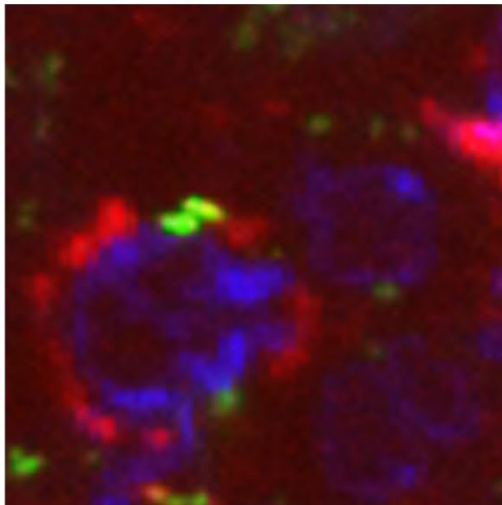
# Correlative Light and Electron Microscopy (CLEM)

1. Taking advantage of both imaging techniques (FL and EM):
  - Specificity of fluorescent markers to identify or pre-select cellular targets
  - High resolution and morphology of EM
2. An efficient approach to the “needle in a haystack” challenge: targeting sparse events in a sample such as proteins, organelles and bacteria.

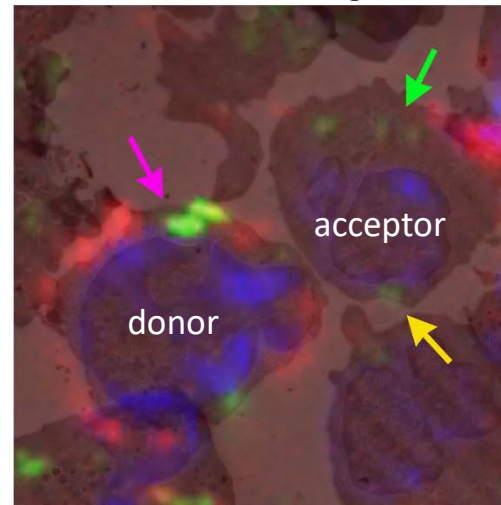
# How to distinguish between sick and healthy cells?

CD45 (donor)  
mitochondria (Dendra2)  
Nuclei (DAPI)

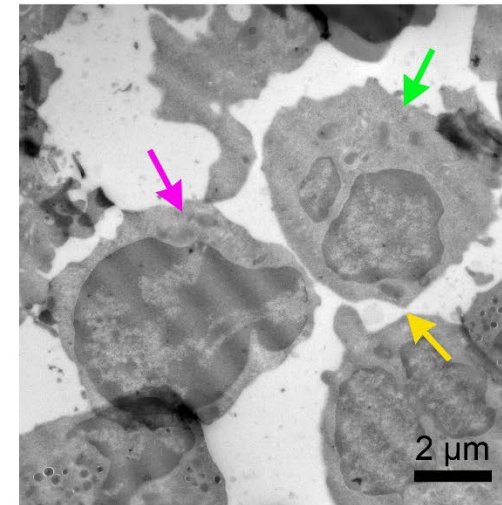
### Fluorescence



### Overlay



### TEM



Donor (healthy) hematopoietic cells transfer functional mitochondria to the irradiated host (sick) bone marrow following total body irradiation.

# Correlative Light and Electron Microscopy (CLEM)

1. Taking advantage of both imaging techniques (FL and EM):
  - Specificity of fluorescent markers to identify or pre-select cellular targets
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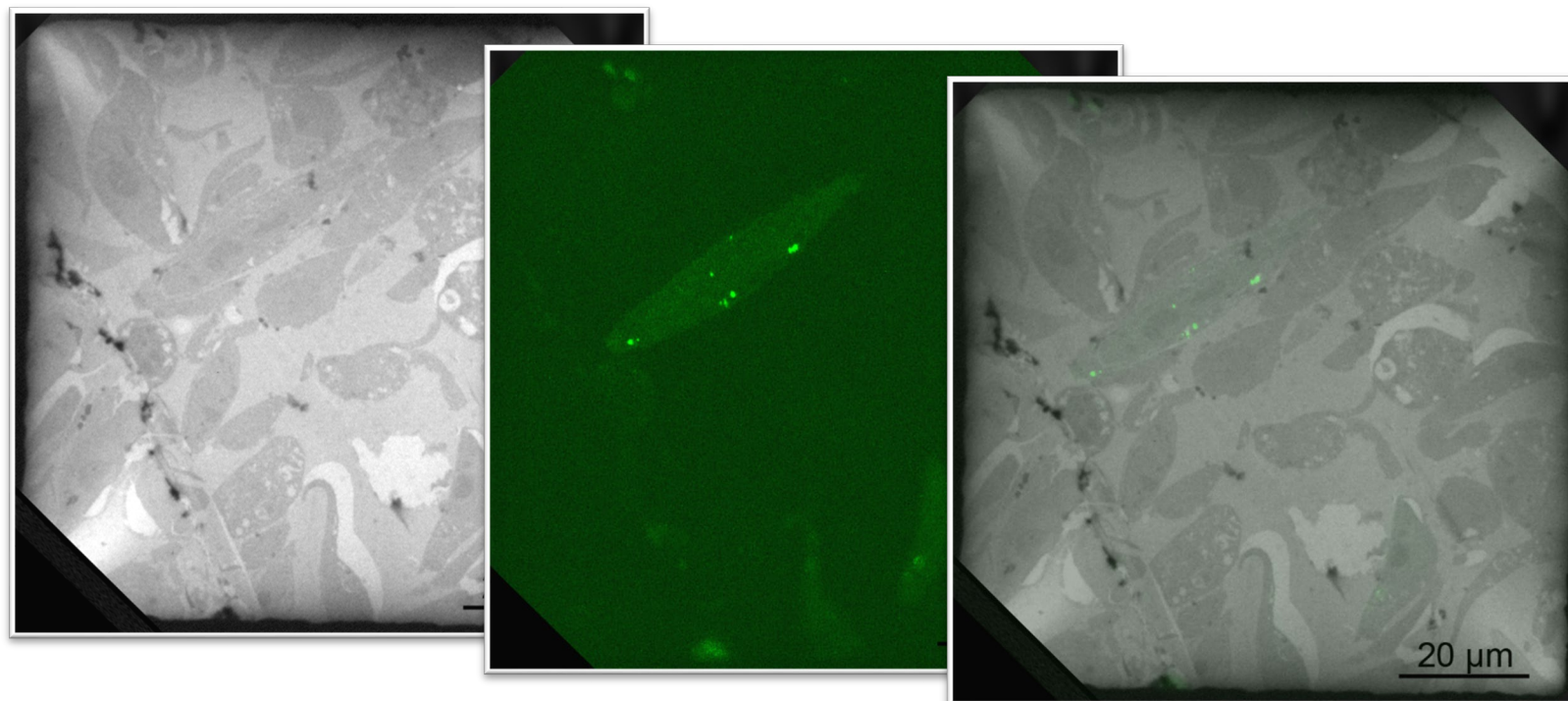
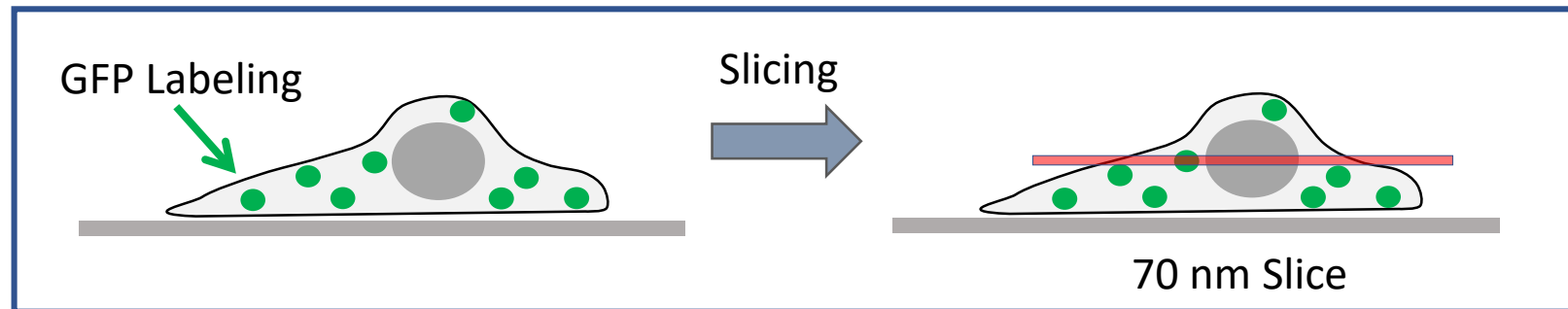
# How long would it take to find Waldo??



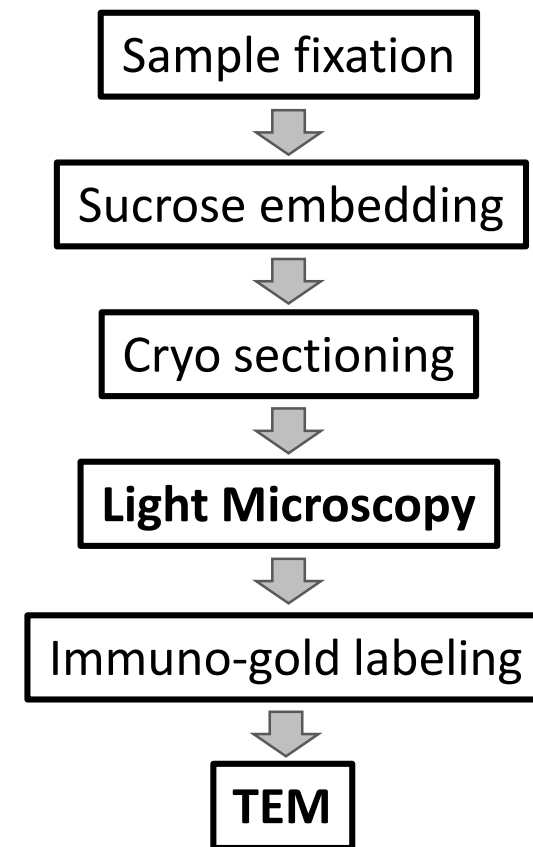
Now...??



# Correlation of Fluorescence and TEM (Tokuyasu technique)

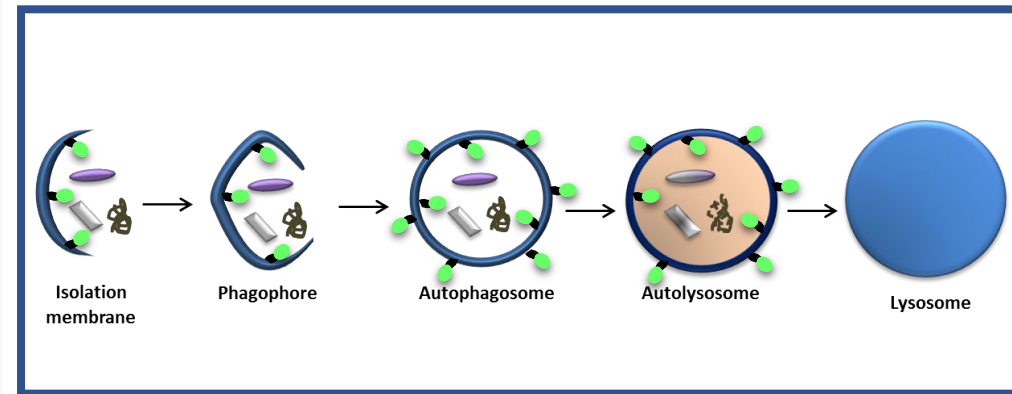


## Correlative workflow

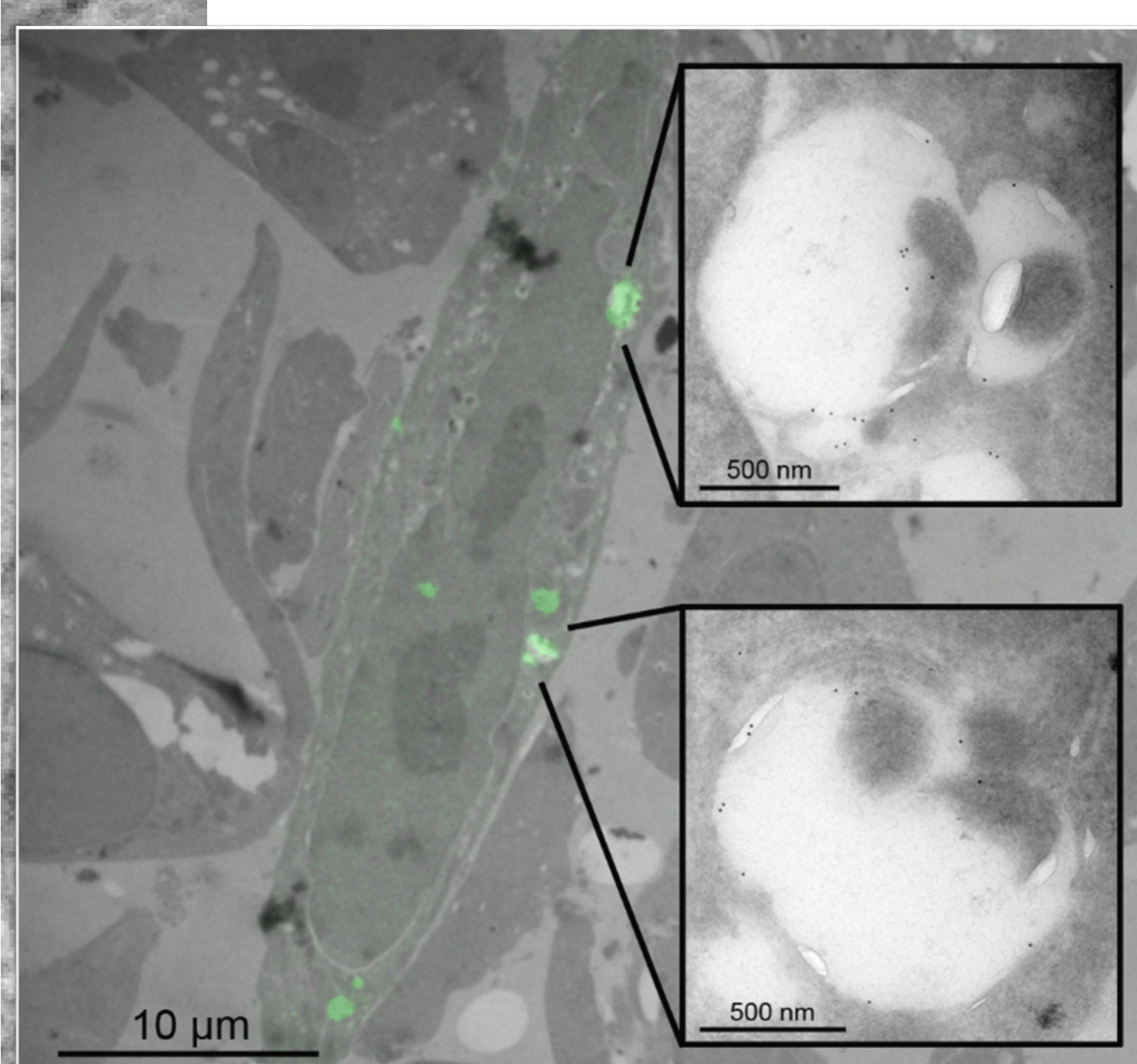




## Autophagosome Biogenesis



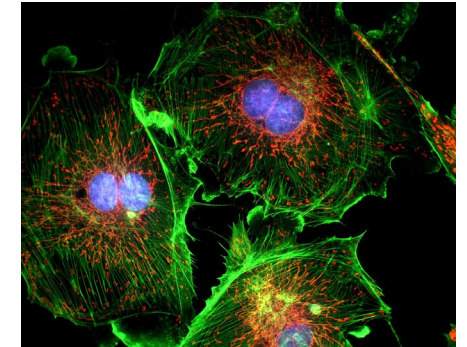
● LC3-GFP-10nm  
Gold NPs



# Challenges

## Fluorescence labeling of the sample

- Genetically encoded fluorescent proteins (GFP, mCherry etc.)
- Organic fluorophores (bright and photo stable).

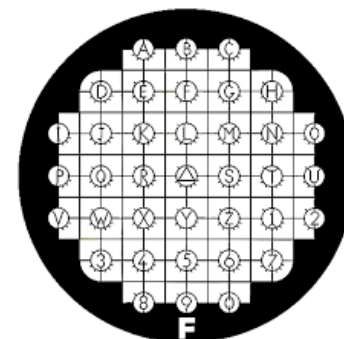


## How to keep the fluorescence alive after treatment for EM?

- Fluorescence is quenched by dehydration, fixatives, heavy metals and resins. Hence, Protocols should be optimized and compromised – specific acrylic resins, no or very little osmium and UrAc...

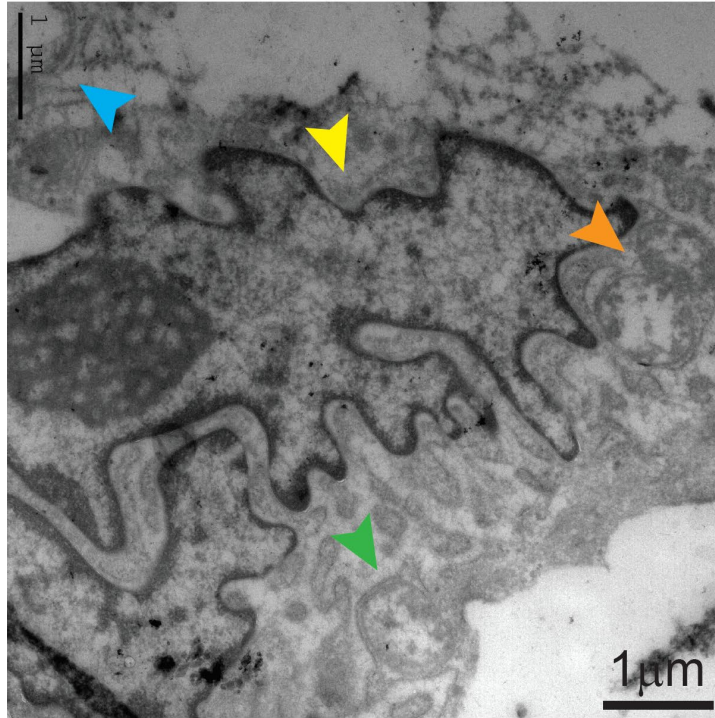
## Registration of images from both modalities

- For navigating and low resolution registration - marked substrate such as finder grid.
- For more precise registration - fiducial markers (FL beads, Nucleus labeling etc.).

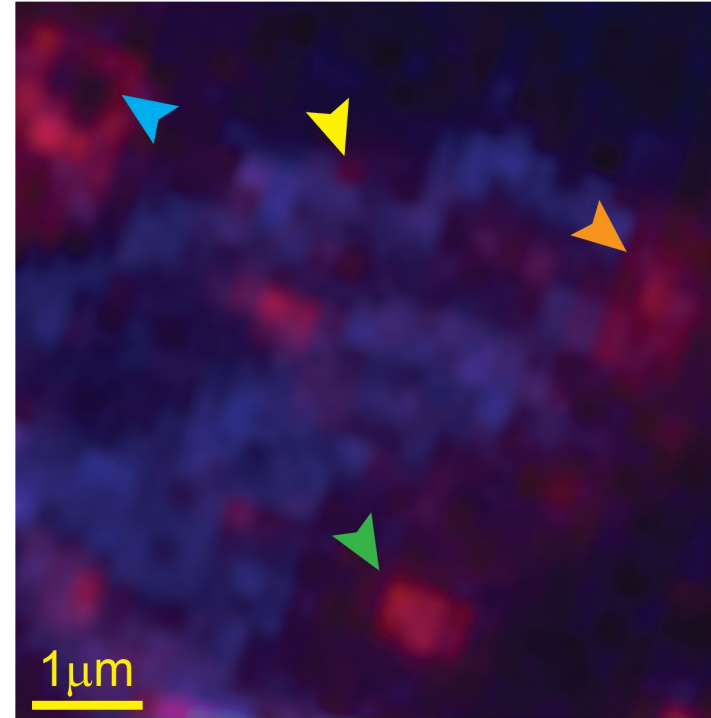


# CLEM targets intra-cellular bacteria in human breast cancer

TEM

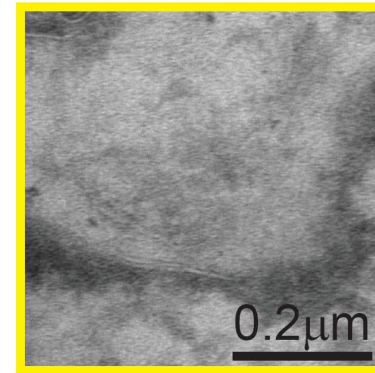
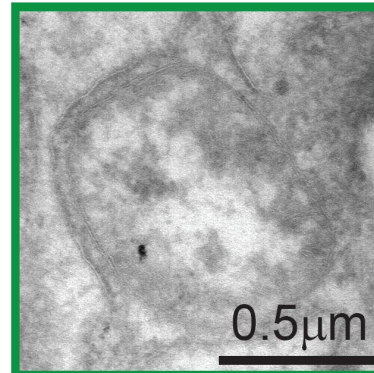
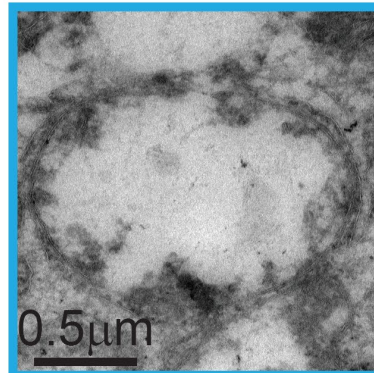
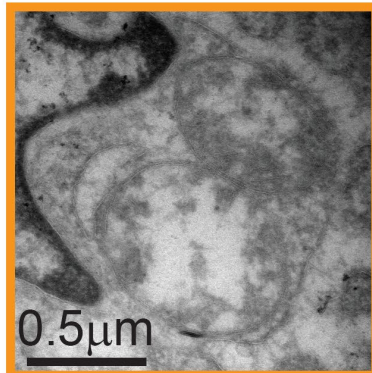


Fluorescence



Identification of bacteria in human breast cancer tumor cells.

Red – Bacteria (Anti-LPS)  
Blue – nucleus (DAPI)



**Dr. Smadar Zaidman**

*Nejman D. et al., Vol. 368, Issue 6494, pp. 973-980, Science 2020*

# Correlative SEM and STORM

The cellulosome (Large multi-enzyme complex) in *Clostridium Clariflavum*

## Correlative workflow

STORM Imaging (fixed sample)



OsO4 post fixation



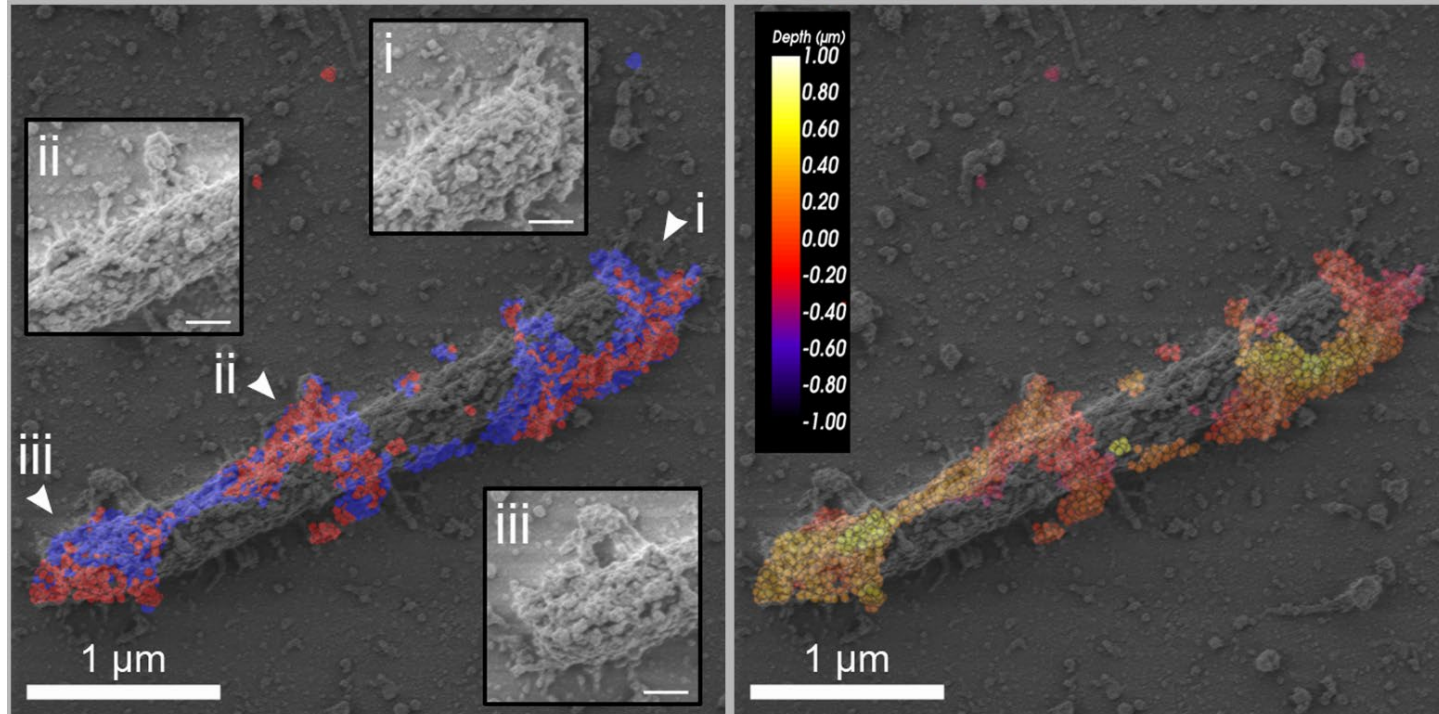
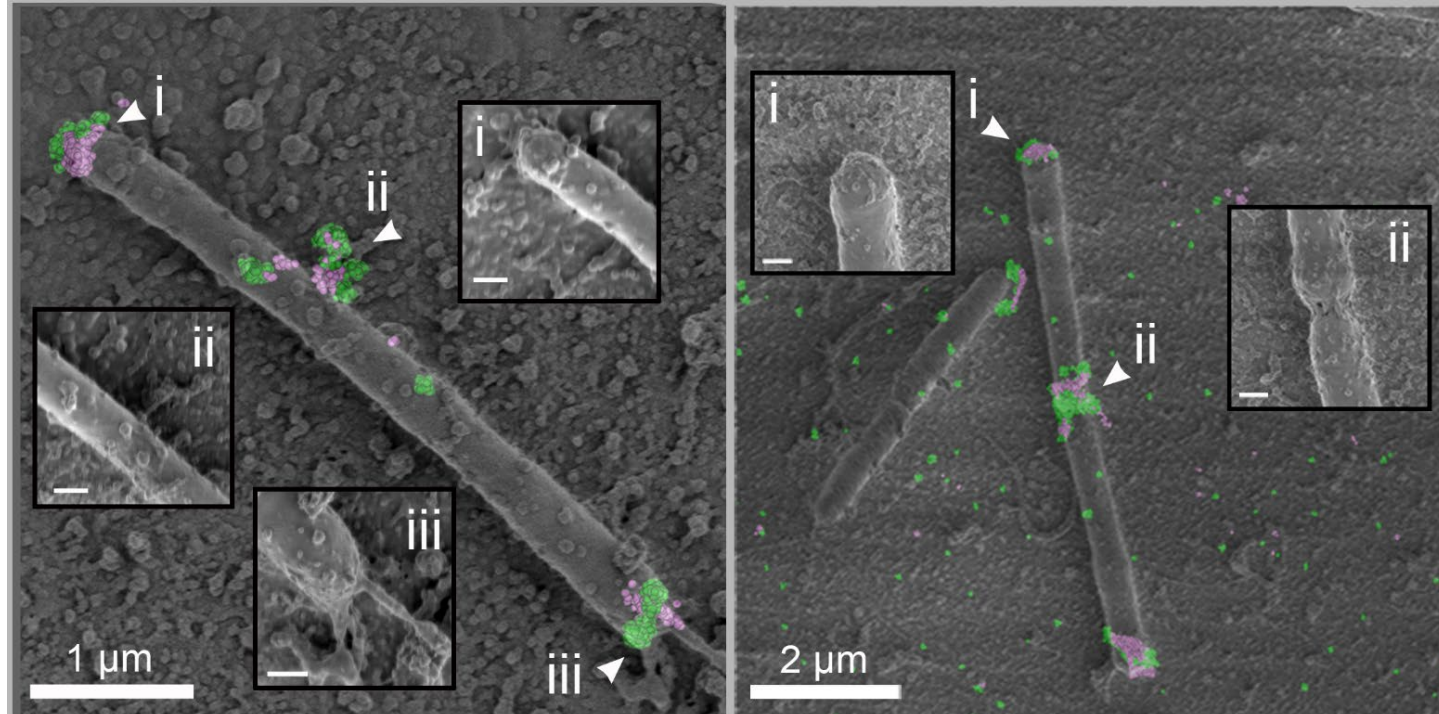
Ethanol dehydration



Critical point drying



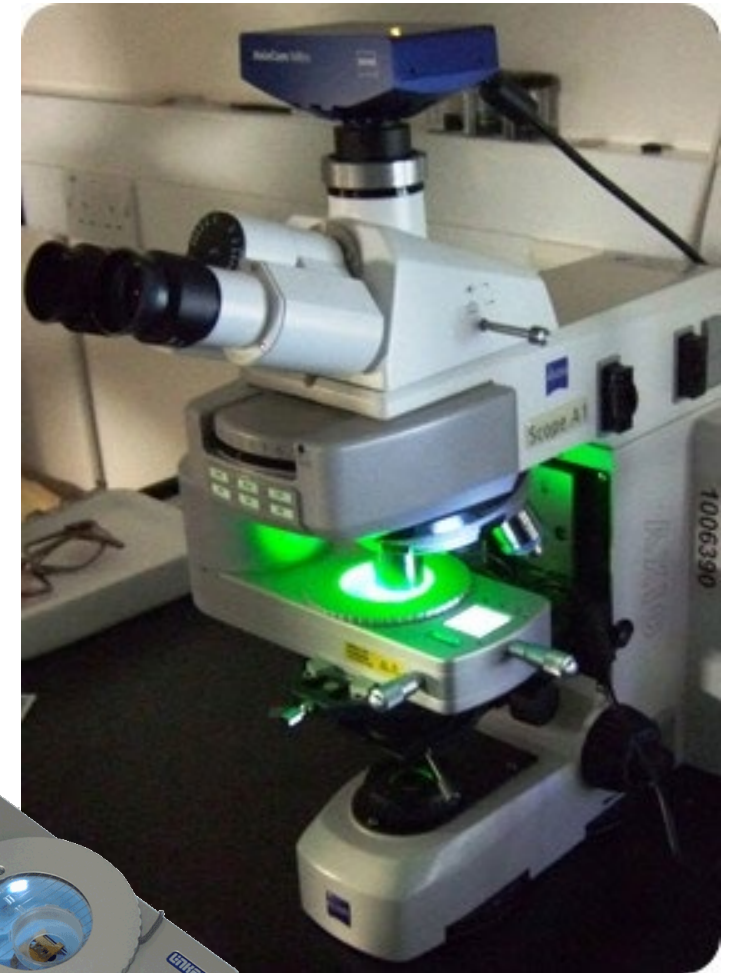
SEM



# Cryo-CLEM

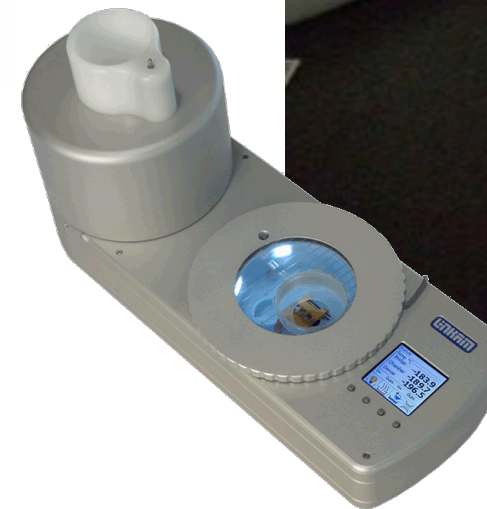


Cryo-CLEM Leica

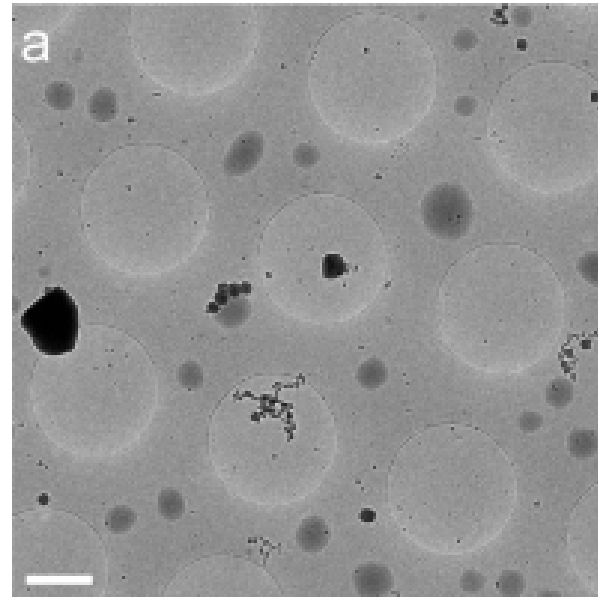


Cryo-CLEM Linkam

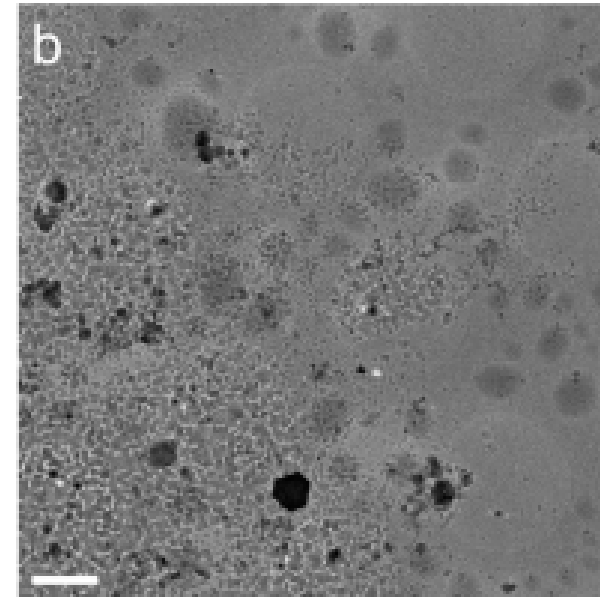
- Stability
- Laser damage
- Long working distance objectives (lower NA)



# Damage of vitreous water by laser illumination



550 W/cm<sup>2</sup>  
30 min



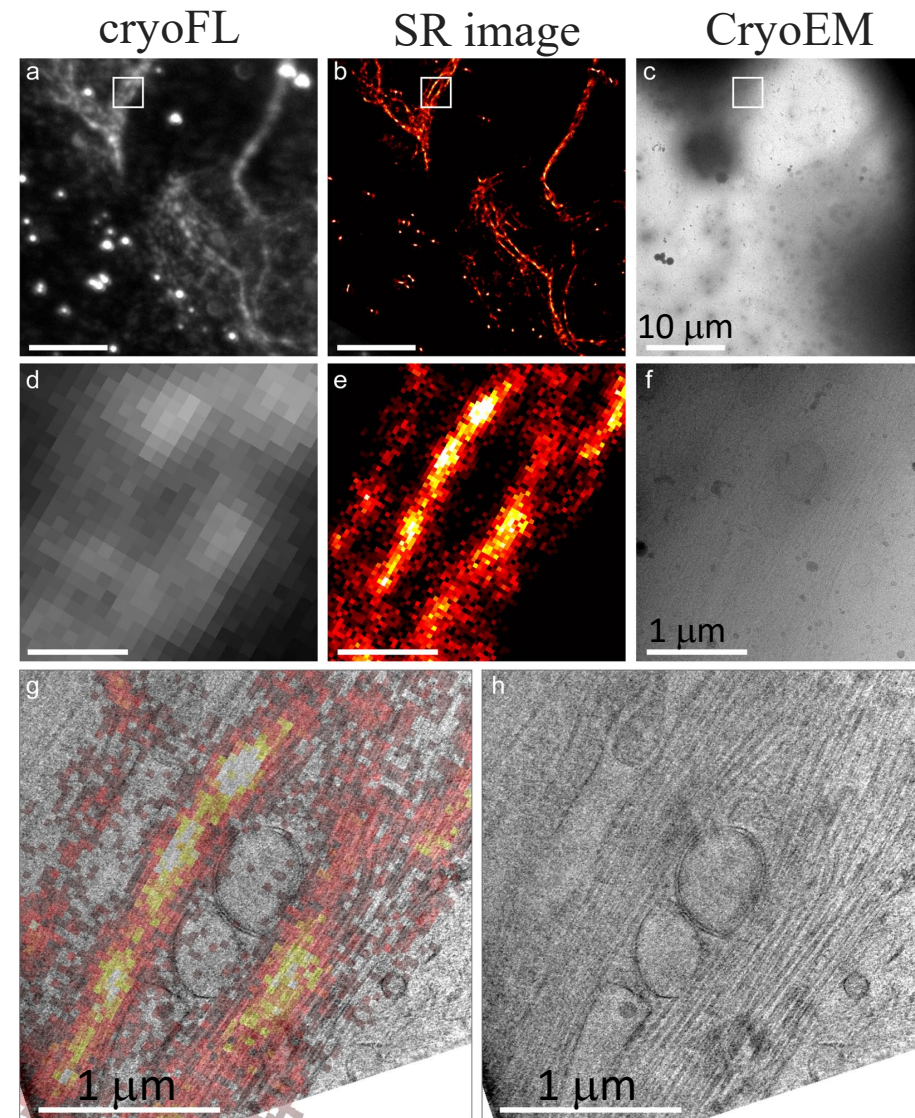
650 W/cm<sup>2</sup>  
5 min

Exposure to high intensity laser light can devitrify cryo samples.

Devitrification is dominated by the laser intensity, and not the illumination time.

# CryoEM and Cryo Super Resolution

Human bone osteosarcoma epithelial (U2OS) cells transfected with plasmid encoding rsEGFP2 fused to microtubule-associated protein 2 (MAP2).



Overlay SR image over 18.6 nm tomographic thick slice

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**Thank you!**

Michael Elbaum - Weizmann